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## (12) United States Patent

Yun et al.

(54) METHOD FOR PREPARING METABOLITES OF ATORVASTATIN USING BACTERIAL CYTOCHROME P450 AND COMPOSITION THEREFOR

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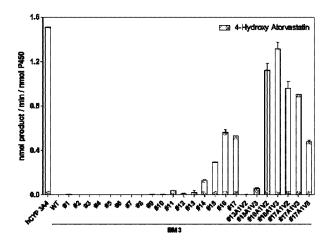
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#### (57) ABSTRACT

Provided is a novel method for preparing metabolites of atorvastatin using bacterial cytochrome P450, and a composition therefor, and more particularly, a composition for preparing 2-hydroxylated product of 4-hydroxylated product from atorvastatin including bacterial cytochrome P 450 BM3 (CYP102A1), CYP102A1 mutants, and chimeras derived from the CYP102A1 mutants, a kit therefor, and a method for preparing thereof.

#### 3 Claims, 14 Drawing Sheets



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Fig. 1

-	
1	MTTKEMPQPKTFGELKNUPLLNTDKPVQALMKTADELGETFKFEAPGRVTRYLSSQRLTK
61	EACDESREDKNESQALKE VROFAGDGLET SWITHEKNWKKABN I LLEPSESQQAMKGYHAMM
121	VDIAVQLVQKWERENADEHTEVPEIMTRETEDTIGLCGFYYRFNSFYRDQPHPFITSMYR
181	ALDEAMNKLQRANPDDPAYDENKRQFQEDIKYMNDLVDKI LADRKASGEQSDDLLTHMLN
241	GKDPETGEPLDDEN IRYQ I ITFL I AGHÆTTSGLLSFÅLYFL VKNPHVLQKA AEEA ARVLV
301	DPVPSYKQVKQLKYVGMYLNEAL8LWPTAPAFSLYAKEDTVLGGEYPLEKGDELMVL1PQ
361	LHRDKTIWGDDVEEFRPERFENPSAIPQHAFKPFGNGQRACIGQQFALHEATLVLGMMLK
421	HEDFEDHTNYELDTKETUTEKPEGEVVKAKSKKTPLGGTPSPSTEQSAKKVRKKÆNAHN
481	TPLLVLYGSAMGTAEGTARDLADIAMSKGFAPQVATLDSHAGNLPREGAVLIVTASYNGH
541	PPDNAKQFVDWLDQASADEVKGVRYSVFGCGDKNWATTYQKVPAFIDETLAAKGAENTAD
601	RGEADASOOFEGTYEEWREHMWSDVAAYFNLDTENSEDNKSTLSLQFYDSAADMPLAKMH
661	GAFSTNYVASKELQQPGSARSTRHLETELPKEASYQEGDHLGVTPRNYEGTVNRYTARPG
721	LDASQQTRLEAEEEKLAHLPLAKTVSVEELLQYVELQDPVTRTQLRAMAAKTVCPPHKVE
781	LEALLEKQAYKEQVLAKRETMLELLEKYPACEMKESEFTALLPSTRPKYYSTSSSPKVDE
841	KQASTTYSVVSGRAWSGYGEYKGTASNYLAELQRGDTTTCFTSTPQSEFTLPKDPETPLT
901	MVGPGTGVAPFRGFVQARKQLKEQGQSLGEAHLYFGCRSPHEDYLYQEELENAQSEG11T
961	LHTAFSRMPNQPKTYVQHVMEQDGKKL FELLDXGAHFY FCGDGSQMAPAVEATLNKSYAD
1021	VHQVSEADARLWLQQLEEKGRYAKDVWAG-

<sup>\*</sup> An amino acid sequence of mutants produced by site-directed mutation of wild-type CYP102Al starts from threonine (T), which is a second amino acid, rather than methionine (M).

Fig. 2

5 - ATGACAATTAAAGAAATGCCTCAGCCAAAAACGTTTGGAGAGCTTAAAAATTTACCGTTATTA AACACAGATAAACCGGTTCAAGCTTTGATGAAAATTGCGGATGAATTAGGAGAAATCTTTAAA TTCGAGGCGCCTGGTGGTAACGCGCTACTTATCAAGTCAGCGTCTAATTAAAGAAGCATGC GATGAATCACGCTTTGATAAAAACTTAAGTCAAGCGCTTAAATTTGTACGTGATTTTGCAGGA GACGGGTTATTTACAAGCTGGACGCATGAAAAAAATTGGAAAAAAGCGCATAATATCTTACTT CCAAGCTTCAGTCAGCAGGCAATGAAAGGCTATCATGCGATGATGGTCGATATCGCCGTGCAG CTTGTTCAAAAGTGGGAGCGTCTAAATGCAGATGAGCATATTGAAGTACCGGAAGACATGACA CGTTTAACGCTTGATACAATTGGTCTTTGCGGCTTTAACTATCGCTTTAACAGCTTTTACCGA GATCAGCCTCATCCATTTATTACAAGTATGGTCCGTGCACTGGATGAAGCAATGAACAAGCTG CAGCGAGCAAATCCAGACGACCCAGCTTATGATGAAAACAAGCGCCAGTTTCAAGAAGATATC  $oldsymbol{a} oldsymbol{a} oldsymbol{G} oldsymbol{a} oldsymbol{G} oldsymbol{G} oldsymbol{G} oldsymbol{A} oldsymbol{G} oldsymbol{G} oldsymbol{A} oldsymbol{A} oldsymbol{A} oldsymbol{A} oldsymbol{G} oldsymbol{G}$ GATGATTTATTAACGCATATGCTAAACGGAAAAGATCCAGAAACGGGTGAGCCGCTTGATGAC GAGAACATTCGCTATCAAATTATTACATTCTTAATTGCGGGACACGAAACAACAAGTGGTCTT TTATCATTTGCGCTGTATTTCTTAGTGAAAAATCCACATGTATTACAAAAAGCAGCAGAAGAA GGCATGGTCTTAAACGAAGCGCTGCGCTTATGGCCAACTGCTCCTGCGTTTTCCCTATATGCA AAAGAAGATACGGTGCTTGGAGGAGAATATCCTTTAGAAAAAGGCGACGAACTAATGGTTCTG ATTCCTCAGCTTCACCGTGATAAAACAATTTGGGGAGACGATGTGGAAGAGTTCCGTCCAGAG CGTTTTGAAAATCCAAGTGCGATTCCGCAGCATGCGTTTAAACCGTTTGGAAACGGTCAGCGT GCGTGTATCGGTCAGCAGTTCGCTCTTCATGAAGCAACGCTGGTACTTGGTATGATGCTAAAA CACTTTGACTTTGAAGATCATACAAACTACGAGCTCGATATTAAAGAAACTTTAACGTTAAAA CCTGAAGGCTTTGTGGTAAAAGCAAAATCGAAAAAATTCCGCTTGGCGGTATTCCTTCACCT AGCACTGAACAGTCTGCTAAAAAAGTACGCAAAAAGGCAGAAAACGCTCATAATACGCCGCTG CTTGTGCTATACGGTTCAAATATGGGAACAGCTGAAGGAACGGCGCGTGATTTAGCAGATATT GCAATGAGCAAAGGATTTGCACCGCAGGTCGCAACGCTTGATTCACACGCCGGAAATCTTCCG CGCGAAGGAGCTGTATTAATTGTAACGGCGTCTTATAACGGTCATCCGCCTGATAACGCAAAG CAATTTGTCGACTGGTTAGACCAAGCGTCTGCTGATGAAGTAAAAGGCGTTCGCTACTCCGTA TTTGGATGCGGCGATAAAAACTGGGCTACTACGTATCAAAAAGTGCCTGCTTTTATCGATGAA ACGCTTGCCGCTAAAGGGGCAGAAAACATCGCTGACCGCGGTGAAGCAGATGCAAGCGACGAC TTTGAAGGCACATATGAAGAATGGCGTGAACATATGTGGAGTGACGTAGCAGCCTACTTTAAC CTCGACATTGAAAACAGTGAAGATAATAAATCTACTCTTTCACTTCAATTTGTCGACAGCGCC CTTCAACAGCCAGGCAGTGCACGAAGCACGCGACATCTTGAAATTGAACTTCCAAAAGAAGCT TOTTATOAAGAAGGAGATOATTTAGGTGTTATTOOTOGOAACTATGAAGGAATAGTAAACOGT GTAACAGCAAGGTTCGGCCTAGATGCATCACAGCAAATCCGTCTGGAAGCAGAAGAAGAAAAA TTAGETCATTTGCCACTCGCTAAAACAGTATCCGTAGAAGAGCTTCTGCAATACGTGGAGCTT CAAGATCCTGTTACGCGCACGCAGCTTCGCGCAATGGCTGCTAAAACGGTCTGCCCGCCGCAT AAAGTAGAGCTTGAAGCCTTGCTTGAAAAGCAAGCCTACAAAGAACAAGTGCTGGCAAAACGT TTAACAATGCTTGAACTGCTTGAAAAATACCCGGCGTGTGAAATGAAATTCAGCGAATTTATC GCCCTTCTGCCAAGCATACGCCCGCGCTATTACTCGATTTCTTCATCACCTCGTGTCGATGAA AAACAAGCAAGCATCACGGTCAGCGTTGTCTCAGGAGAAGCGTGGAGCGGATATGGAGAATAT AAAGGAATIGCGTCGAACTATCTTGCCGAGCTGCAAGAAGGAGATACGATTACGTGCTTIATT  ${\tt TCCACACCGCAGTCAGAATTTACGCTGCCAAAAGACCCTGAAACGCCGCTTATCATGGTCGGA$ CCGGGAACAGGCGTCGCGCGTTTAGAGGCTTTGTGCAGGCGCGCAAACAGCTAAAAGAACAA GGACAGTCACTTGGAGAAGCACATTTATACTTCGGCTGCCGTTCACCTCATGAAGACTATCTG TATCAAGAAGAGCTIGAAAACGCCCAAAGCGAAGGCATCATTACGCTTCATACCGCTTTTTCT CGCATGCCAAATCAGCCGAAAACATACGTTCAGCACGTAATGGAACAAGACGGCAAGAAATTG ATTGAACTTCTTGATCAAGGAGCGCACTTCTATATTTGCGGAGACGGAAGCCAAATGGCACCT CGCTTATGGCTGCAGCAGCTAGAAGAAAAAGGCCGATACGCAAAAGACGTGTGGGCTGGGTAA-31

Fig. 3

Amino acid sequence of wild-type CYP102Al mutant #16 (M16)

.1	MITTREMPQPKTFGELKNLPLLNTDRPVQALMKTADELGETPRFEAPGRVTRYLSSQRLTR
61	EACDESRFDKNLSQALKFYRDFAGIXELFTSWTHEKNWKKAHN LLLPSFSQQAMKGYHAMM
121	VDTAVQLVQKWERENADEHTEVPEDMTRETEDTTGLCGPNYRENSFYRDQPHPFTTSMVR
181	ALDEAMNKLQRANPDDPAYDENKRQEQED IKVMNDL VDK I I ADRKASGEQSDDLL TIMLN
241	GKDPETGEPLDDEN I RYQ I I TTL I AGHETTSGLLSFALYFLVKNPHVLQKAAEEAARVLV
301	DPVPSYKQVKQLKYVGNVLNEALRLWPTAPAFSLYAKEDTVLGGEYPLEKGDELVVLIPQ
361	LHRDKT I WGDDVEEFRPERFENPSA I PQHAFKPFGNGQRAC I GQQFALHEATL VLGMMLK
421	HPDFEDHTNYELDIKETLTLKPEGFVVKAKSKKIPLGGIPSPSTEQSAKKVRKKAENAHN
481	TPLLVLYGSNMGTAEGTARDLADIAMSKGFAPQVATLDSHAGNLPREGAVLIVTASYNGH
541	PPDNAKQFVDWLDQASADEVKGVRYSVFGCGDKNWATTYQKVPAFIDETLAAKGAENIAD
601	RGEADASDDFEGTYEEWREHMWSDVAAYFNLDTENSEDNKSTUSUQFVDSAADMPLAKMH
661	GAFSTNVVASKELQQPGSARSTRIILE I ELPKEASYQEGDIILGV I PRNYEG I VNRVTARFG
721	LDASQQTRLEAEEEKLAHUPLAKTYSVEELLQYVELQOPVTRTQLRAMAAKTVCPP9KVE
781	LEALLEKQAYKEQVLAKRITMLELLEKYPACEMKESEFTALLPSTRPRYYSTSSSPRVDE
841	KQASITVSVVSGEAWSGYGEYKGIASNYLAÆLQEGDTITCPISTPQSEFTLPKDFÆTPLI
901	MVGPGTGVAPFRGFVQARKQLKEQGQSLGEAHLYFGCRSPHEDYLYQEELENAQSEGTIT
961	LHTAFSRMPNQPKTYVQHVMEQDGKKL1ELLDQGAHFYTCGDGSQMAPAVEATLWKSYAD
1021	VIQVSEADARLWLQQLEEKGRYAKDVWAG-

#### Fig. 4

5' -ATGACAATTAAAGAAATGCCTCAGCCAAAAACGTTTGGAGAGCTTAAAAATTTACCGTTATTA AACACAGATAAACGGGTTCAAGCTTTGATGAAAATTGGGGATGAATTAGGAGAAATCTTTAAA TTCGAGGCGCCTGGTCTTGTAACGCGCTACTTATCAAGTCAGCGTCTAATTAAAGAAGCATGC GATGAATCACGCTTTGATAAAAACTTAAGTCAAGCGCTTAAATTTGTACGTGATATTGCAGGA GACGGGTTAGTTACAAGCTGGACGCATGAAAAAATTGGAAAAAAGCGCATAATATCTTACTT CCAAGCTTCAGTCAGCAGGCAATGAAAGGCTATCATGCGATGATGGTCGATATCGCCGTGCAG CTTGTTCAAAAGTGGGAGCGTCTAAATGCAGATGAGCATATTGAAGTACCGGGAGACATGACA CGTTTAACGCTTGATACAATTGGTCTTTGCGGCTTTAACTATCGCTTTAACAGCTTTTACCGA GATCAGCCTCATCCATTTATTACAAGTATGGTCCGTGCACTGGATGAAGCAATGAACAAGCAG CAGCGAGCAAATCCAGACGACCCAGCTTATGATGAAAACAAGCGCCAGTTTCAAGAAGATATC GATGATTTATTAACGCATATGCTAAACGGAAAAGATCCAGAAACGGGTGAGCCGCTTGATGAC GAGAACATICGCIAICAAATTATTACATICTTAATTGCGGGACACGTAACAACAAGTGGTCTI TTATCATTTGCGCTGTATTTCTTAGTGAAAAATCCACATGTATTACAAAAAGCAGCAGAAGAA GGCATGGTCTTAAACGAAGCGCTGCGCTTATGGCCAACTGCTCCTGCGTTTTCCCTATATGCA AAAGAAGATACGGTGCTTGGAGGAGAATATCCTTTAGAAAAAGGCGACGAACTAATGGTTCTG ATTCCTCAGCTTCACCGTGATAAAACAATTTGGGGAGACGATGTGGAAGAGTTCCGTCCAGAG CGTTTTGAAAATCCAAGTGCGATTCCGCAGEATGCGTTTAAACCGTTTGGAAACGGTCAGCGT GCGTGTATCGGTCAGCAGTTCGCTCTTCATGAAGCAACGCTGGTACTTGGTATGATGCTAAAA CACTTTGACTTTGAAGATCATACAAACTACGAGCTCGATATTAAAGAAACTTTAACGTTAAAA CCTGAAGGCTTTGTGGTAAAAGCAAAATCGAAAAAATTCCGCTTGGCGGTATTCCTTCACCT AGCACTGAACAGTCTGCTAAAAAAGTACGCAAAAAGGCAGAAAACGCTCATAATACGCCGCTG CTTGTGCTATACGGTTCAAATATGGGAACAGCTGAAGGAACGGCGCGTGATTTAGCAGATATT GCAATGAGCAAAGGATTTGCACCGCAGGTCGCAACGCTTGATTCACACGCCGGAAATCTTCCG CGCGAAGGAGCTGTATTAATTGTAACGGCGTCTTATAACGGTCATCCGCCTGATAACGCAAAG CANTTIGICGACTOGITAGACCAAGCGICIGCTGATGAAGTAAAAGGCGITCGCTACTCCGIA TTTGGATGCGGCGATAAAAACTGGGCTACTACGTATCAAAAAGTGCCTGCTTTTATCGATGAA ACGCTTGCCGCTAAAGGGGCAGAAAACATCGCTGACCGCGGTGAAGCAGATGCAAGCGACGAC TTTGAAGGCACATATGAAGAATGGCGTGAACATATGTGGAGTGACGTAGCAGCCTACTTTAAC CTCGACATTGAAAACAGTGAAGATAATAAATCTACTCTTTCACTTCAATTTGTCGACAGCGCC CTTCASCAGCCAGGCAGTGCACGAAGCACGCGACATCTTGAAATTGAACTTCCAAAAGAAGCT TOTTATOAAGAAGGAGATOATITAGGTGTTATTOOTOGCAACTATGAAGGAATAGTAAACOGT GTAACAGCAAGGTTCGGCCTAGATGCATCACAGCAAATCCGTCTGGAAGCAGAAGAAGAAAAA TTAGCTCATTTGCCACTCGCTAAAACAGTATCCGTAGAAGAGCTTCTGCAATACGTGGAGCTT CAAGATCCTGTTACGCGCACGCAGCTTCGCGCAATGGCTGCTAAAACGGTCTGCCCGCCGCAT AAAGTAGAGCTTGAAGCCTTGCTTGAAAAGCAAGCCTACAAAGAACAAGTGCTGGCAAAACGT TTAACAATGCTTGAACTGCTTGAAAAATACCCGGCGTGTGAAATGAAATTCAGCGAATTTATC GCCCTTCTGCCAAGCATACGCCCGCGCTATTACTCGATTTCTTCATCACCTCGTGTCGATGAA AAACAAGCAAGCATCACGGTCAGCGTTGTCTCAGGAGAAGCGTGGAGCGGATATGGAGAATATA A A G G A A T I G C G T C G A A C T A T C T T G C C G A G C T G C A A G A A G G A G A T A C G A T C A C G T G C T T T A T T TCCACACCGCAGTCAGAATTTACGCTGCCAAAAGACCCTGAAACGCCGCTTATCATGGTCGGA  $\texttt{CCGGGAACAGGCGTCGCGCCGTTTAGAGGCTTTGTGCAGGCGCGCAAACAGCTAAAAGAACAA$ GGACAGTCACTTGGAGAAGCACATTTATACTTCGGCTGCCGTTCACCTCATGAAGACTATCTG TATCAAGAAGAGCTTGAAAACGCCCAAAGCGAAGGCATCATTACGCTTCATACCGCTTTTTCT CGCATGCCAAATCAGCCGAAAACATACGTTCAGCACGTAATGGAACAAGACGGCAAGAAATTG ATTGAACTTCTTGATCAAGGAGCGCACTTCTATATTTGCGGGAGACGGAAGCCAAATGGCACCT CGCTTATGGCTGCAGCAGCTAGAAGAAAAAGGCCGATACGCAAAAGACGTGTGGGCTGGGTAA-3'

Fig. 5 Amino acid sequence of wild-type CYP102A1 mutant #17 (M17)

1	MT I KEMPQPKTFGELKNLPLLNTDXPVQALMK I ADELGE I FKFEAPGLVTRYLSSQRL I K
61	EACDGSRFDKNLSQALKFVRDTAGDGLVTSWTHEKNWKKAIDNTLLPSFSQQAMKGYHAMM
121	VDIAVQLVQKWERLAADEHIEVPGDMTRLTLDTIGLCGFNYRFNSFYRDQPHPFITSMVR
181	ALDEAMNKQQRANPODPAYDENKRQFQEDIKVMNDLVDKII ADRKASGEQSDDLLTHMLN
241	GKDPETGEPLDDEN FRYQT I TFL I AGHVTTSGLLSFALYFLVKNPHVLQKAAEEAARVLV
301	DPVPSYKQVKQLKYVGMVLNEALREWPTAPAFSLYAKEDTVLGGEYPLEKGDELMVLIPQ
361	LHRDKT I WGDDVEEFRPERFENPSA I PQHAFKPFGNGQRAC I GQQFALHEATL VLGMMLK
421	HFDFEDITTNYELD I KETLTLKPEGFVVKAKSKK I PLGG I PSPSTEQSAKKVRKKVENAHN
481	TPLLVLYGSNMGTAEGTARDLAD I AMSKGPAPQVATLDSHAGNLPREGAVLIVTASYNGH
541	PPDNAKQFVDWLDQASADDVKGVRYSVFGCGDKNWATTYQKVPAF1DETLAAKGAEN1AD
601	RGEADASODFEGTYEEWREHMWSDVAAYFNLDTENSEDNESTLSLQFVDSAADMPLAKME
661	GAFSANVVASKELQQLGSERSTRHLETALPKEASYQEGDHLGVTPRNYEGTVNRVTARFG
721	LDASQQ IRLEAEEEKLAHLPLGKTVSVEELLQYVELQDPVTRTQLRAMAAKTVCPPHKVE
781	LEALLEKQAYKEQVLAKRETMLELLEKYPACEMEFSEFIALLPSISPRYYSISSSPHVDE
841	KQASITVSVVSGEAWSGYGEYKGIASNYLANLQEGDTITCFVSTPQSGFTLPKDSETPLI
901	MVGPGTGVAPFRGFVQARKQLKEQGQSLGEAHLYFGCRSPHEDYLYQEELENAQNEG11T
961	LHTAFSRVPNQPKTYVQHVMERDGKKLIELLDQGAHFYTCGDGSQMAPDVEATLMKSYAD
1021	VYEVSEADARLWLQQLEEKGRYAKDVWAG-

#### Fig. 6

5. -ATGACAATTAAAGAAATGCCTCAGCCAAAAACGTTTGGAGAGCTTAAAAATTTACCGTTATTA AACACAGATAAACGGGTTCAAGCTTTGATGAAAATTGCGGATGAATTAGGAGAAATCTTTAAA TTCGAGGCGCCTGGTCTTGTAACGCGCTACTTATCAAGTCAGCGTCTAATTAAAGAAGCATGC GATGGATCACGCTTTGATAAAAACTTAAGTCAAGCGCTTAAATTTGTACGTGATATTGCAGGA GACGGGTTAGTTACAAGCTGGACGCATGAAAAAATTGGAAAAAAGCGCATAATATCTTACTT CCAAGCITCAGTCAGCAGGCAATGAAAGGCTATCATGCGATGATGGTCGATATCGCCGTGCAG CTTGTTCAAAAGTGGGAGCGTCTAAATGCAGATGAGCATATTGAAGTACCGGGAGACATGACA CGTTTAACGCTTGATACAAITGGICITIGCGGCTTTAACTATCGCTITAACAGCTTTTACCGA GATCAGCCTCATCCATTTATTACAAGTATGGTCCGTGCACTGGATGAAGCAATGAACAAGCAG CAGCGAGCAAATCCAGACGACCCAGCTTATGATGAAAACAAGCGCCAGTTTCAAGAAGATATC A A G G T G A T G A A C G A C C T A G T A G A T A A A A T T A T T G C A G A T C G C A A A G C G A G C G T G A A C A A A G C GATGATITATIAACGCATATGCTAAACGGAAAAGATCCAGAAACGGGTGAGCCGCTTGATGAC GAGAACATTCGCTATCAAATTATTACATTCTTAATTGCGGGACACGTAACAACAAGTGGTCTT TTATCATITGCGCIGIAITTCTTAGTGAAAAATCCACAIGIAITACAAAAAGCAGCAGAAGAA GCAGCACGAGITCTAGIAGATCCTGITCCAAGCTACAAACAAGTCAAACAGCTTAAATATGTC GGCATGGTCTTAAACGAAGCGCTGCGCTTATGGCCAACTGCTCCTGCGTTTTCCCCTATATGCA A A A G A A G A T A C G G T G C T I G G A G G A G A A T A T C C T T I A G A A A A A G G C G A C G A A C T A A T G G T T C T G ATTCCTCAGCTTCACCGTGATAAAACAATTTGGGGAGACGATGTGGAAGAGTTCCGTCCAGAG CGTTTTGAAAATCCAAGTGCGATTCCGCAGCATGCGTTTAAACCGTTTGGAAACGGTCAGCGT GCGTGTATCGGTCAGCAGTTCGCTCTTCATGAAGCAACGCTGGTACTTGGTATGATGCTAA4A CACTTTGACTTTGAAGATCATACAAACTACGAGCTCGATATTAAAGAAACTTTAACGTTAAAA CCTGAAGGCTTTGTGGTAAAAGCAAAATCGAAAAAATTCCGCTTGGCGGTATTCCTTCACCT AGUACT GAACAGT CTGCTAAAAAAGTAGGCAAAAAGGCAGAAAACGCTCATAATACGCCGCTG CTTGTGCTATACGGTTCAAATATGGGAACAGCTGAAGGAACGGCGCGTGATTTAGCAGATATT GCAATGAGCAAAGGATTTGCACCGCAGGTCGCAACGCTTGATTCACACGCCGGAAATCTTCCG CGCGAAGGAGCTGTATTAATTGTAACGGCGTCTTATAACGGTCATCCGCCTGATAACGCAA4G CAATTTGTCGACTGGTTAGACCAAGCGTCTGCTGATGAAGTAAAAGGCGTTCGCTACTCCGTA TTTGGATGCGGCGATAAAAACTGGGCTACTACGTATCAAAAAGTGCCTGCTTTTATCGATGAA ACGCTTGCCGCTAAAGGGGCAGAAAACATCGCTGACCGCGGTGAAGCAGATGCAAGCGACGAC TTTGAAGGCACATATGAAGAATGGCGTGAACATATGTGGAGTGACGTAGCAGCCTACTTTAAC OTOGACATIGAAAACAGTGAAGATAATAAATOTACICITICACITOAATITGTOGACAGOGCO CTTCAACAGCCAGGCAGTGCACGAAGCACGCGACATCTTGAAATTGAACTTCCAAAAGAAGCT TOTTATOAAGAAGGAGATCATTTAGGTGTTATTOOTOGCAACTATGAAGGAATAGTAAACOGT GTAACAGCAAGGTTCGGCCTAGATGCATCACAGCAAATCCGTCTGGAAGCAGAAGAAGAAAAA TTAGCTCATTTGCCACTCGCTAAAACAGTATCCGTAGAAGAGCTTCTGCAATACGTGGAGCTT CAAGATCCIGITACGCGCACGCAGCTTCGCGCAATGGCTGCIAAAACGGTCTGCCCGCCGCAT AAAGTAGAGCTTGAAGCCTTGCTTGAAAAGCAAGCCTACAAAGAACAAGTGCTGGCAAAACGT TTAACAATGCTTGAACTGCTTGAAAAATACCCGGGGTGTGAAATGAAATTCAGCGAATTTATC GCCCTTCTGCCAAGCATACGCCCGCGCTATTACTCGATTTCTTCATCACCTCGTGTCGATGAA AAACAAGCAAGCATCACGGTCAGCGTTGTCTCAGGAGAGCGTGGAGCGGATATGGAGAATAT AAAGGAATTGCGTCGAACTATCTTGCCGAGCTGCAAGAAGGAGATACGATTACGTGCTTTATT TCCACACCGCAGICAGAATTTACGCTGCCAAAAGACCCTGAAACGCCGCTTATCATGGTCGGA CCGGGAACAGGCGTCGCGCCGTTTAGAGGCTTTGTGCAGGCGCGCAAACAGCTAAAAGAACAA GGACAGTCACTTGGAGAAGCACATTTATACTTCGGCTGCCGTTCACCTCATGAAGACTATCTG TATCAAGAAGAAGCTTGAAAACGCCCAAAGCGAAGGCATCATTACGCTTCATACCGCTTTTTCT C G C A T G C C A A T C A G C C G A A A A C A T A C G T T C A G C A C G T A A T G G A A C A A G A C G G C A A G A A T T G ATTGAACTTCTIGATCAAGGAGCGCACTTCTATATTTGCGGAGACGGAAAGCCAAATGGCACCT CGCTTATGGCTGCAGCAGCTAGAAGAAAAAGGCCGATACGCAAAAGACGTGTGGGCTGGGTAA-3:

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Fig. 7

# Amino acid sequence of chimera M16A1V2 derived from wild-type CYP102A1 mutant #16 (M16)

1	MTTKEMPQPKTFGELKNLPLLNTDKPVQALMKTADELGETFRFEAPGRYTRYLSSQRLTK
61	EACDESRFTWNLSQALKFVRDFAGDGLFTSWTHEKNWKKAANTLLPSFSQQAMKGYHAMM
121	VDIAVQLVQKWERLNADEHIEVPEDMTRLTLDTIGLCGFNYRFNSFYRDQPHPFITSMVR
181	ALDEAMNKLQRANPODPAYDENKRQFQEDIKVMNDEVDKITADRKASGEQSODLLTHMEN
241	GKDPETGEPLDDENTRYQTTTFLTAGHETTSGLLSFALYFLVKNPHVLQKAAEEAARVLV
301	DPVPSYKQVKQLKYVGMVLNEALRINPTAPAFSLYAKEDTVLGGEYPLEKGDELMVL (PQ
361	LHRDKTTWGDDVEEFRPERFENPSATPQHAFKPFGNGQRACTGQQFALHEATLVLGMMLK
421	HFDFEDHTNYELDIKETLTLKPEGFVVKAKSKKIPLGGIPSPSTEQSAKKVRKK <b>VE</b> NAHN
481	TPLLVEVGSNMGTÆRTÆRÐEAÐFAMSKGFÆPQVATEÐSHAGNEPREGAVELIVTASYNGIF
541	PPDNAKQFVDWLDQASADDVKGVRYSVFGCGDKNWATTYQKVPAFIDETLAAKGAENTAD
601	RGEADASDDFEGTYEEWREIMWSDVAAYFNLDIENSEDAKSTLSLQFVDSAADMPLAKMII
661	GAFSANVVASKELQQLGSERSTRHLE FALPKEASYQEGDHLGV I PRNYEG I VNRVTAKFG
721	LDASQQIRLEAEEEKLAHLPLGKTVSVEELLQYVELQDPVTRTQLRAMAAKTVCPPHKVE
781	LEALLEKQAYKEQVLAKRLTMLELLEKYPACEMEFSEFTALLPSTSPRYYSTSSSPHVDE
841	KQASTTVSVVSGEAWSGYGEYKGTASNYLANLQEGDTTTCFVSTPQSGFTLPKDSETPLT
901	MVGPGTGVAPFRGFVQARKQLKEQGQSLGEAHLYFGCRSPHEDYLYQEELENAQNEG LIT
961	LHTAFSRVPNQPKTYVQHVMERDGKKLHELLDQGA9FYTCGDGSQMAPDVEATLMKSYAD
1021	VYEVSEADARLWLQQLEEKGRYAKDVWAG-

Fig. 8

5 - ATGACAATTAAAGAAATGCCTCAGCCAAAAACGTTTGGAGAGCTTAAAAATTTACCGTTATTA AACACAGATAAACCGGTTCAAGCTTTGATGAAAATTGCGGATGAATTAGGAGAAATCTTTAAA TTCGAGGCGCCTGGTCTTGTAACGCGCTACTTATCAAGTCAGCGTCTAATTAAAGAAGCATGC GATGAATCACGCTTTGATAAAACTTAAGTCAAGCGCTTAAATTTGTACGTGATATTGCAGGA GACGGGTTAGTTACAAGCTGGACGCATGAAAAAATTGGAAAAAAGCGCATAATATCTTACTT CCAAGETTCAGTCAGCAGGCAATGAAAGGCTATCATGCGATGATGGTCGATATCGCCGTGCAG CTTGTTCAAAAGTGGGAGCGTCTAAATGCAGATGAGCATATTGAAGTACCG**GGA**GACATGACA CGTTTAACGCTTGATACAATTGGTCTTTGCGGCTTTAACTATCGCTTTAACAGCTTTTACCGA GATCAGCCTCATCCATTTATIACAAGTATGGTCCGTGCACTGGATGAAGCAATGAACAAGCAG CAGUGAGCAAATCCAGACGACUCAGCTTATGATGAAAACAAGCGCCAGTTTCAAGAAGATATC GATGATTTATTAACGCATATGCTAAACGGAAAAGATCCAGAAACGGGTGAGCCGCTTGATGAC GAGAACATICGCIATCAAATTATTACATTCTTAATIGCGGGACACGTAACAACAAGTGGICTT TTATCATTTGCGCTGTATTTCTTAGTGAAAAATCCACATGTATTACAAAAAGCAGCAGAAGAA GCAGCACGAGTTCFAGTAGATCCTGTTCCAAGCTACAAACAAGTCAAACAGCTTAAATATGTC GGCATGGTCTTAAACGAAGCGCTGCGCTTATGGCCAACTGCTCCTGCGTTTTCCCTATATGCA AAAGAAGATACGGTGCTTGGAGGAGAATATCCTTTAGAAAAAGGCGACGAACTAATGGTTCTG ATTCCTCAGCTTCACCGTGATAAAACAATTTGGGGAGACGATGTGGAAGAGTTCCGTCCAGAG CGTTTTGAAAATCCAAGTGCGATTCCGCAGCATGCGTTTAAACCGTTTGGAAACGGTCAGCGT GCGTGTATCGGTCAGCAGTTCGCTCTTCATGAAGCAACGCTGGTACTTGGTATGATGCTAAAA CACITIGACITIGAAGATCATACAAACTACGAGCICGATAITAAAGAAACITTAACGTTAAAA CCTGAAGGCTTTGTGGTAAAAGCAAAATCGAAAAAATTCCGCTTGGCGGTATTCCTTCACCT AGCACTGAACAGTCIGCTAAAAAAGTACGCAAAAAGGTAGAAAACGCICATAATACGCCGCIG CTTGTGCTATACGGTTCAAATATGGGAACAGCTGAAGGAACGGCGCGTGATTTAGCAGATATT GCAATGAGCAAAGGATTTGCACCGCAGGTCGCAACGCTTGATTCACACGCCGGAAATCTTCCG CGCGAAGGAGCTGTATIAATIGTAACGGCGTCTTATAACGGTCATCCGCCTGATAACGCAAAG CAATTTGTCGACTGGTTAGACCAAGCGTCTGCTGATGATGTAAAAGGCGTTCGCTACTCCGTA TTTGGATGCGGCGATAAAAACTGGGCTACTACGTATCAAAAAGTGCCTGCTTTTATCGATGAA ACGCTTGCCGCTAAAGGGGCAGAAAACATCGCTGACCGCGGTGAAGCAGATGCAAGCGACGAC TTTGAAGGCACATATGAAGAATGGCGTGAACATATGTGGAGTGACGTAGCAGCCTACTTTAAC CTCGACATTGAAAACAGTGAAGATAATAAATCTACTCTTTCACTTCAATTTGTCGACAGCGCC GCGGATATGCCGCTTGCGAAAATGCACGGTGCGTTTTCAGCGAACGTCGTAGCAAGGAAAGAA CTTCAACAGCTAGGCAGTGAACGAAGCACGCGACATCTTGAAATTGCACTTCCAAAAGAAGCT TCTTATEAAGAAGGAGATEATTTAGGTGTTATTEETCGCAACTATGAAGGAATAGTAAACCGT GTAACAGCAAGGTTCGGCCTAGATGCATCACAGCAAATCCGTCTGGAAGCAGAAGAAGAAAAA TTAGCTCATTTGCCACTCGGTAAAACAGTATCCGTAGAAGAGCTTCTGCAATACGIGGAGCTT CAAGATOCTGTTACGCGCACGCAGCTTCGCGCAATGGCTGCTAAAACGGTCTGCCCGCCGCAT AAAGTAGAGCTTGAAGCCTTGCTTGAAAAGCAAGCCTACAAAGAACAAGTGCTGGCAAAAAGT TTAACAATGCTTGAACTGCTTGAAAAATACCCGGCGTGTGAAATGGAATICAGCGAATTTATC GCCCTTCTGCCAAGCATAAGCCCGCGCTATTACTCGATTTCTTCATCACCTCATGTCGATGAA AAACAAGCAAGCATCACGGTCAGCGTTGTCTCAGGAGAGAGCGTGGAGCGGATATGGAGAGATAT AAAGGAATIGCGICGAACIAICTTGCCGATCTGCAAGAAGGAGATACGATTACGTGCITTGTT TCCACACCGCAGTCAGGATTTACGCTGCCAAAAGACTCTGAAACGCCGCTTATCATGGTCGGA CCGGGAACAGGCGICGCGCGTTTAGAGGCTTIGIGCAGGCGCGCAAACAGCTAAAAGAACAA GGACAGTCACTTGGAGAAGCACATTTATACTTCGGCTGCCGTTCACCTCATGAAGACTATCTG TATCAAGAAGAGOTTGAAAACGCCCAAAACGAAGGCAICATTACGCTTCATACCGCTTTTTCT CGCGTGCCAAATCAGCCGAAAACATACGITCAGCACGTAAIGGAACGAGACGGCAAGAAAITG ATTGAACTTCTTGATCAAGGAGCGCACTTCTATATTTGCGGAAGGCGAAGCCAAATGGCACCT GACGTTGAAGCAACGCTTATGAAAAGCIA1GCTGACGITTACGAAGTGAGTGAGGAGACGCT CGCTTATGGCTGCAGCAGCTAGAAGAAAAAGGCCGATACGCAAAAGACGTGTGGGCTGGGTAA~3'

Sep. 8, 2015

Fig. 9

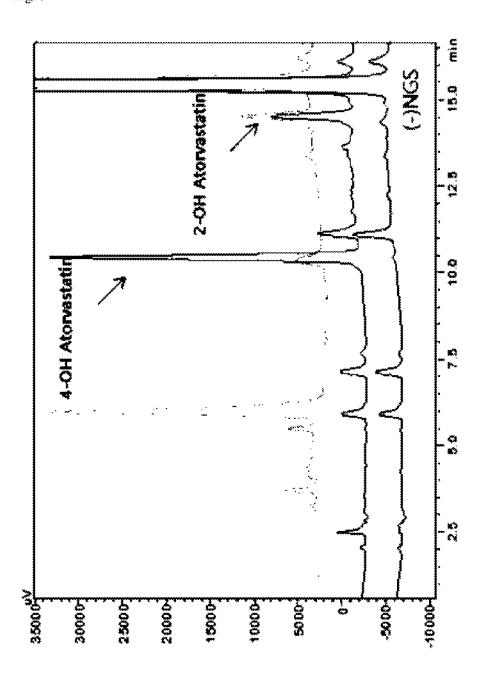
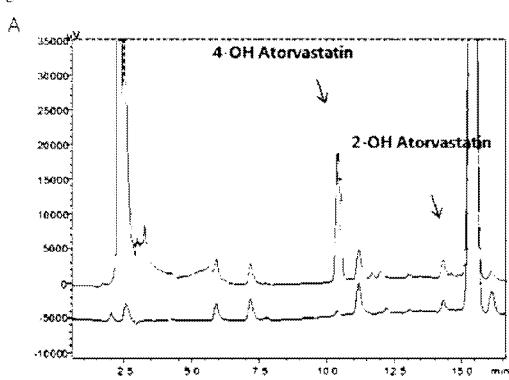
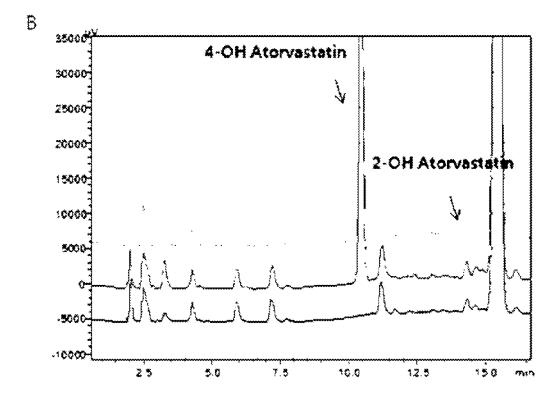
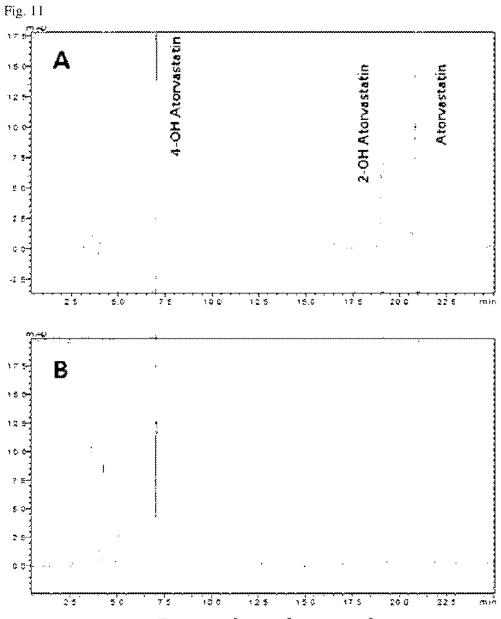


Fig. 10

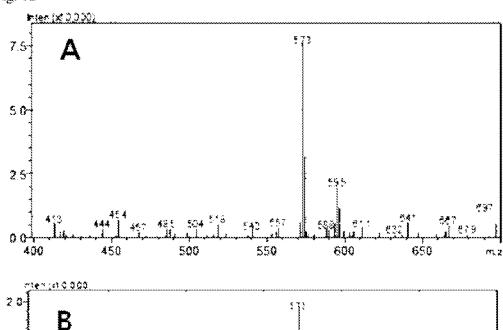


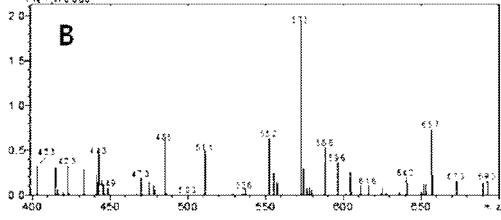




Retention time, min

Fig. 12





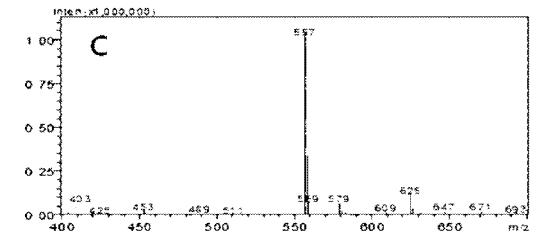


Fig. 13

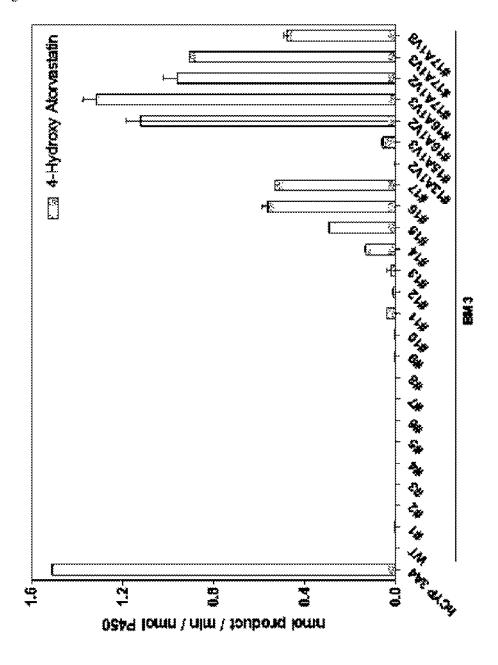
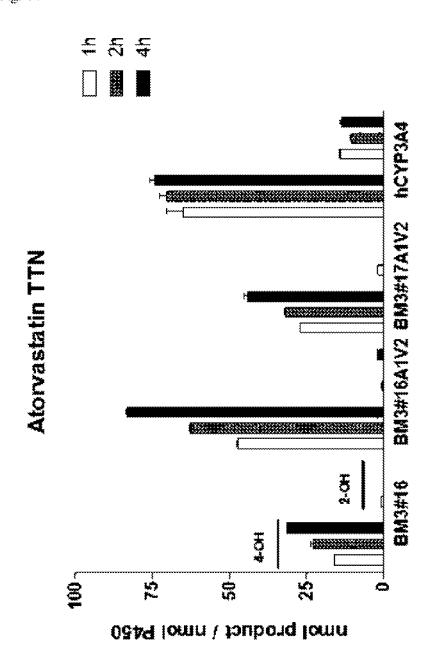


Fig. 14



#### METHOD FOR PREPARING METABOLITES OF ATORVASTATIN USING BACTERIAL CYTOCHROME P450 AND COMPOSITION THEREFOR

#### TECHNICAL FIELD

The present invention relates to a novel method for preparing metabolites of atorvastatin using bacterial cytochrome P450 and a composition therefor.

#### **BACKGROUND ART**

Atorvastatin is well known as an anti-hyperlipidemic agent, an antihypercholesterolemic agent, or a cholesterol- 15 lowering agent. Oxidative metabolism of atorvastatin in human liver is mediated by mainly cytochrome P450 3A (CYP3A) enzymes, particularly, cytochrome P450 3A4 (CYP3A4), and the following two metabolites, that is, orthoastatin) and parahydroxy atorvastatin (para-OH atorvastatin or 4-OH atorvastatin) are generated.

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HMG-CoA reductase catalyzes the conversion of HMG-CoA to mevalonate, which is an early and rate-limiting step in the biosynthesis of cholesterol.

In addition to the P450-mediated oxidation and β-oxidation processes, glucuronidation constitutes a common metabolic pathway for statins (Prueksaritanont et al., Drug Metab. Dispos. 30:505-512, 2002). The metabolites resulting from microsomal oxidation of atorvastatin by P450 enzymes are effective inhibitors of HMG-CoA reductase. In addition, it has been suggested that the metabolites may contribute to the cholesterol-lowering effect of atorvastatin.

Cytochrome P450 enzymes (P450s or CYPs) are large families consisting of enzymes serving as remarkably diverse oxygenation catalysts in throughout nature from archaea, bacteria, fungi, plants, and animals up to humans (http:// drnelson.uthsc.edu/CytochromeP450.html). Due to the catalytic diversity and broad substrate range of P450s, they are attractive biocatalyst candidates for the production of fine chemicals, including pharmaceuticals.

However, in spite of the potential use of mammalian P450shydroxy atorvastatin (ortho-OH atorvastatin or 2-OH atorv- 20 in various biotechnology fields, they are not suitable as biocatalysts because of their low stability, low catalytic activity, and low affordability.

After oral ingestion, atorvastatin, which is an inactive lacform. This is a main metabolite and an inhibitor of 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase.

In the case in which a pro-drug is converted into a biologitone, is hydrolyzed to the corresponding β-hydroxy acid 65 cally "active metabolite" by human hepatic P450s during drug development, a large amount of pure metabolites are required in order to research into effect, toxicity, pharmaco-

kinetics of the drug, or the like. Further, in the case in which the metabolite itself has biological activity, it may be advantageous to directly administer the metabolite to the body. Therefore, it is important to prepare the metabolite on a large scale

However, since there are various problems in chemically synthesizing pure metabolites, P450 may be used in order to prepare the metabolites of a drug or drug candidates as an alternative for chemical synthesis of the metabolites. The metabolite preparation has been reported using human P450s expressed in *Escherichia coli* (Yun et al., Curr. Drug Metab. 7:411-429, 2006) and in insect cells (Rushmore et al., Metab. Eng. 2:115-125, 2000; Vail et al., J. Ind. Microbiol. Biotechnol. 32:67-74. 2005).

However, since these systems are still costly and have low productivities due to limited stabilities and slow reaction rates, a method of using engineered bacterial P450 enzymes having the desired catalyst activity has been suggested as an alternative for producing human metabolite.

Meanwhile, P450 BM3 (CYP102A1) from *Bacillus megaterium* has strong similarity to eukaryotic members of the CYP4A (fatty acid hydroxylase) family. It has been reported that CYP102A1 mutants oxidizes several human P450 substrates to produce the metabolite with higher activity (Kim et al., Protein Expr. Purif. 57:188-200, 2008a). Further, CYP102A1 is a versatile monooxygenase capable of working on various substrates (Di Nardo et al., J. Biol. Inorg. Chem. 12:313-323, 2007).

Recently, it has been reported that CYP102A1 mutants may produce larger quantities of the human metabolites of drugs, which may be difficult to be synthesized (Otey et al., Biotechnol. Bioeng. 93:494-499, 2005). Therefore, as an alternative method of preparing the metabolites, it may be considered to use CYP102A1 engineered so as to have the desired properties.

Several amino acid residues in CYP102A1 were mutated to generate mutant enzymes having increased activity toward human P450 substrates by the present inventors (Yun et al., 40 Trends Biotechnol. 25:289-298, 2007 and other references cited in the article), and it was confirmed that specific mutants among these mutant enzymes may enable the CYP102A1 enzyme to catalyze O-deethylation and 3-hydroxylation of 7-ethoxycoumarin (Kim et al. Drug Metab. Dispos. 36:2166-45 2170, 2008a).

Therefore, while conducting research for directly using the atorvastatin metabolites as a drug, the present inventors discovered bacterial enzymes capable of oxidizing atorvastatin, which is known as a human P450 substrate, to produce 2-hydroxylated product and 4-hydroxylated product, which are human metabolites, and a biological preparation method using the same, thereby completing the present invention.

#### DISCLOSURE OF INVENTION

#### Technical Problem

An object of the present invention is to provide a bacterial enzyme capable of oxidizing atorvastatin to preparing 4-hydroxylated product or 2-hydroxylated product, which are human metabolites, on a large scale.

In addition, another object of the present invention is to provide a composition for preparing 2-hydroxylated product or 4-hydroxylated product from atorvastatin containing the enzyme. 4

Further, another object of the present invention is to provide a method for preparing 2-hydroxylated product or 4-hydroxylated product from atorvastatin including reacting the enzyme with atorvastatin.

Furthermore, another object of the present invention is to provide a kit for preparing 2-hydroxylated product or 4-hydroxylated product from atorvastatin containing the enzyme and a reduced nicotinamide adenine dinucleotide phosphate (NADPH)-generating system.

#### Solution to Problem

In one general aspect, there is provided a preparation method capable of selectively preparing human metabolites, particularly 2-hydroxylated product or 4-hydroxylated product from atorvastatin on a large scale using wild-type CYP102A1, CYP102A1 mutants, or chimeras derived from CYP102A1 mutants as a bacterial P450 enzyme, and a composition and a kit therefor.

In the present invention, "the CYP102A1 mutants" have an amino acid sequence of the wild-type CYP102A1 modified by natural or artificial substitution, deletion, addition, and/or insertion. Preferably, amino acid of the CYP102A1 mutant may be substituted with an amino acid that has similar properties as classified below. For example, alanine, valine, leucine, isoleucine, proline, methionine, phenylalanine, and tryptophan are classified as nonpolar amino acids and have similar properties to each other. Glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine are neutral amino acids, aspartic acid and glutamic acid are acidic amino acids, and lysine, arginine, and histidine are basic amino acids.

The CYP102A1 mutants according to the present invention include polypeptide having an amino acid sequence similar to an amino acid sequence of CYP102A1 at an identity level of 50% or more, preferably, 75% or more, and more preferably, 90% or more.

In the present invention, the terms "chimeric" is used in the case in which at least two binding domains that are different from each other are contained therein. The two binding domains may be derived from different wild-type proteins. The two domains may be derived from the same wild-type protein, but in chimeric protein according to the present invention, the two domains may be positioned in a different arrangement from the corresponding the wild-type CYP102A1 mutant protein by fusing a heme domain of the wild-type CYP102A1 and a reductase domain of natural variants of the wild-type CYP102A1 to each other.

Hereinafter, the present invention will be described in detail.

The wild-type CYP102A1, the CYP102A1 mutant, or the chimera derived from the CYP102A1 mutant may be used as a catalyst in oxidation reaction using atorvastatin that is known as a human P450 substrate as the substrate.

More specifically, the present inventors clarified that the wild-type CYP102A1, the CYP102A1 mutant, or the chimera derived from the CYP102A1 mutant may be used as a catalyst in oxidation reaction using atorvastatin that is known as a human P450 substrate as the substrate. Particularly, in the case in which human CYP3A4 is used as the catalyst, as the produced atorvastatin metabolites, 2-hydroxylated product and 4-hydroxylated product may not be selectively produced. On the other hand, in the case in which the wild-type CYP102A1 mutant and the chimeras derived from the CYP102A1 according to the present invention are used as the catalyst, large amounts of 2-hydroxylated product and 4-hydroxylated product may be selectively and stably produced.

The present inventors prepared chimeras (#16A1V2, #17A1V2) derived from the CYP102A1 by selecting several mutants (wild-type CYP102A1 mutants #16 and #17 shown in Tables 2 and 3) with high catalytic activity for some substrates in a human among mutants prepared by over-expressing bacterial wild-type CYP102A1 and site-directed mutants thereof in *E. coli* (See Table 1) and fusing heme domains thereof and reductase domains of natural variants of the wild-type CYP102A1 to each other.

In the case in which the bacterial wild-type CYP102A1, the 10 prepared mutants thereof (wild-type CYP102A1 mutants #16 and #17 shown in Tables 2 and 3), and chimeras (#16A1V2, #17A1V2) derived from the CYP102A1 was over-expressed in *E. coli* to be reacted with atorvastatin and a NADPH-generating system, it was confirmed that atorvastatin is converted into metabolites in humans through high-performance liquid chromatography (HPLC) (See FIG. 9) and a liquid chromatography-mass spectrometry (LC-MS) spectrum (See FIGS. 11 and 12).

In the case in which human CYP3A4 is used as the catalyst, 20 as the produced atorvastatin metabolites, 2-hydroxylated product and 4-hydroxylated product may not be selectively produced. On the other hand, it might be appreciated that in the case in which the wild-type CYP102A1 mutant and the chimeras derived from the CYP102A1 according to the 25 present invention are used as the catalyst, 2-hydroxylated product and 4-hydroxylated product may be selectively prepared on a large scale.

In addition, it might be appreciated that three kinds of mutants (#15, #16, and #17 in Table 2) and five kinds of 30 chimeras (#16A1V2, #16A1V3, #17A1V2, #17A1V3, and #17A1V8) derived from the mutants have a large turnover number among the wild-type CYP102A1 mutants and the chimeras derived from the wild-type CYP102A1 mutants in producing the metabolites of atorvastatin. Particularly, it 35 might be appreciated that the chimera #16A1V2 derived from the CYP102A1 mutant #16 and the chimera #17A1V2 derived from the CYP102A1 mutant #17 have the most excellent turnover number. See FIG. 14.

Based on the experiment results as described above, the 40 present invention provides a composition for preparing 2-hydroxylated product or 4-hydroxylated product from atorvastatin including at least one enzyme selected from a group consisting of the wild-type CYP102A1, the CYP102A1 mutants, and chimeras derived from the CYP102A1 mutants, 45

wherein the CYP102A1 mutant has an amino acid sequence changed from that of the wild-type CYP102A1 by at least one substitution selected from a group consisting of substituting arginine (R) at the amino acid position 47 with an amino acid selected from a group consisting of alanine, 50 valine, leucine, isoleucine, proline, methionine, phenylalanine, and tryptophan, substituting tyrosine (Y) at the amino acid position 51 with an amino acid selected from a group consisting of alanine, valine, isoleucine, proline, methionine, phenylalanine, and tryptophan, substituting glutamic acid (E) 55 at the amino acid position 64 with an amino acid selected from a group consisting of glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine, substituting alanine (A) at the amino acid position 74 with an amino acid selected from a group consisting of glycine, serine, threonine, 60 cysteine, tyrosine, asparagine, and glutamine, substituting phenylalanine (F) at the amino acid position 81 with an amino acid selected from a group consisting of alanine, valine, leucine, isoleucine, proline, methionine, and tryptophan, substituting leucine (L) at the amino acid position 86 with an amino acid selected from a group consisting of alanine, valine, isoleucine, proline, methionine, phenylalanine, and tryptophan,

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substituting phenylalanine (F) at amino acid position 87 with an amino acid selected from a group consisting of alanine, valine, leucine, isoleucine, proline, methionine, and tryptophan, substituting glutamic acid (E) at the amino acid position 143 with an amino acid selected from a group consisting of glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine, substituting leucine (L) at the amino acid position 188 with an amino acid selected from a group consisting of glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine, and substituting glutamic acid (E) at the amino acid position 267 with an amino acid selected from a group consisting of alanine, valine, leucine, isoleucine, proline, methionine, phenylalanine, and tryptophan, and

the chimera derived from the CYP102A1 mutant has an amino acid sequence changed from that of the reductase domain of the CYP102A1 mutant by at least one substitution selected from a group of substituting lysine (K) at the amino acid position 474 with threonine (T), substituting alanine (A) at the amino acid position 475 with valine (V), substituting glutamine (O) at the amino acid position 513 with arginine (R), substituting arginine (R) at the amino acid position 526 with proline (P), substituting glutamine (Q) at the amino acid position 547 with glutamic acid (E), substituting glutamic acid (E) at the amino acid position 559 with aspartic acid (D), substituting leucine (L) at the amino acid position 590 with phenylalanine (F), substituting alanine (A) at the amino acid position 591 with serine (S), substituting aspartic acid (D) at the amino acid position 600 with glutamic acid (E), substituting valine (V) at the amino acid position 625 with leucine (L), substituting aspartic acid (D) at the amino acid position 632 with asparagine (N), substituting aspartic acid (D) at the amino acid position 638 with glutamic acid (E), substituting lysine (K) at the amino acid position 640 with alanine (A), substituting alanine (A) at the amino acid position 652 with serine (S), substituting glycine (G) at the amino acid position 661 with arginine (R), substituting threonine (T) at the amino acid position 665 with alanine (A), substituting glutamine (Q) at the amino acid position 675 with lysine (K), substituting proline (P) at the amino acid position 676 with leucine (L), substituting alanine (A) at the amino acid position 679 with glutamic acid, substituting glutamic acid (E) at the amino acid position 688 with alanine (A), substituting threonine (T) at the amino acid position 716 with alanine (A), substituting alanine (A) at the amino acid position 717 with threonine (T), substituting alanine (A) at the amino acid position 742 with glycine (G), substituting alanine (A) at the amino acid position 783 with valine (V), substituting alanine (A) at the amino acid position 796 with threonine (T), substituting lysine (K) at the amino acid position 814 with glutamic acid (E), substituting isoleucine (I) at the amino acid position 825 with methionine (M), substituting arginine (R) at the amino acid position 826 with serine (S), substituting arginine (R) at the amino acid position 837 with histidine (H), substituting glutamic acid (E) at the amino acid position 871 with asparagine (N), substituting isoleucine (I) at the amino acid position 882 with valine (V), substituting glutamic acid (E) at the amino acid position 888 with glycine (G), substituting aspartic acid (D) at the amino acid position 894 with glycine (G), substituting proline (P) at the amino acid position 895 with serine (S), substituting glycine (G) at the amino acid position 913 with serine (S), substituting glutamic acid (E) at the amino acid position 948 with lysine (K), substituting serine (S) at the amino acid position 955 with asparagine (N), substituting methionine (M) at the amino acid position 968 with valine (V), substituting glutamine (Q) at the amino acid position 971 with glutamic acid (E), substituting methionine (M) at the amino acid position 980 with valine (V), substituting

glutamine (Q) at the amino acid position 982 with arginine (R), substituting alanine (A) at the amino acid position 1009 with aspartic acid (D), substituting aspartic acid (D) at the amino acid position 1020 with glutamic acid (E), substituting histidine (H) at the amino acid position 1022 with tyrosine 5 (Y), substituting glutamine (Q) at the amino acid position 1023 with lysine (K) and glutamic acid (E), and substituting glycine (G) at the amino acid position 1040 with serine (S).

Further, in another general aspect, the present invention provides a method for preparing 2-hydroxylated product or 10 4-hydroxylated product from atorvastatin including reacting at least one enzyme selected from a group consisting of wild-type CYP102A1, CYP102A1 mutants, and chimeras derived from the CYP102A1 mutants with atorvastatin,

wherein the CYP102A1 mutant has an amino acid 15 sequence changed from that of the wild-type CYP102A1 by at least one substitution selected from a group consisting of substituting arginine (R) at the amino acid position 47 with an amino acid selected from a group consisting of alanine, valine, leucine, isoleucine, proline, methionine, phenylala- 20 nine, and tryptophan, substituting tyrosine (Y) at the amino acid position 51 with an amino acid selected from a group consisting of alanine, valine, isoleucine, proline, methionine, phenylalanine, and tryptophan, substituting glutamic acid (E) at the amino acid position 64 with an amino acid selected 25 from a group consisting of glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine, substituting alanine (A) at the amino acid position 74 with an amino acid selected from a group consisting of glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine, substituting phenylalanine (F) at the amino acid position 81 with an amino acid is selected from a group consisting of alanine, valine, leucine, isoleucine, proline, methionine, and tryptophan, substituting leucine (L) at the amino acid position 86 with an amino acid selected from the group consisting of alanine, 35 valine, isoleucine, proline, methionine, phenylalanine, and tryptophan, substituting phenylalanine (F) at the amino acid position 87 with an amino acid selected from a group consisting of alanine, valine, leucine, isoleucine, proline, methionine, and tryptophan, substituting glutamic acid (E) at 40 the amino acid position 143 with an amino acid selected from a group consisting of glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine, substituting leucine (L) with the amino acid position 188 with an amino acid selected from a group consisting of glycine, serine, threonine, cys- 45 teine, tyrosine, asparagine, and glutamine, and substituting glutamic acid (E) at the amino acid position 267 with an amino acid selected from a group consisting of alanine, valine, an leucine, isoleucine, proline, methionine, phenylalanine, and tryptophan, and

the chimera derived from the CYP102A1 mutant has an amino acid sequence changed from that of the reductase domain of the CYP102A1 mutant by at least one substitution selected from a group of substituting lysine (K) at the amino acid position 474 of the of CYP102A1 mutant with threonine 55 (T), substituting alanine (A) at the amino acid position 475 with valine (V), substituting glutamine (Q) at the amino acid position 513 with arginine (R), substituting arginine (R) at the amino acid position 526 with proline (P), substituting glutamine (Q) at the amino acid position 547 with glutamic 60 acid (E), substituting glutamic acid (E) at the amino acid position 559 with aspartic acid (D), substituting leucine (L) at the amino acid position 590 with phenylalanine (F), substituting alanine (A) at the amino acid position 591 with serine (S), substituting aspartic acid (D) at the amino acid position 65 600 with glutamic acid (E), substituting valine (V) at the amino acid position 625 with leucine (L), substituting aspar8

tic acid (D) at the amino acid position 632 with asparagine (N), substituting aspartic acid (D) at the amino acid position 638 with glutamic acid (E), substituting lysine (K) at the amino acid position 640 with alanine (A), substituting alanine (A) at the amino acid position 652 with serine (S), substituting glycine (G) at the amino acid position 661 with arginine (R), substituting threonine (T) at the amino acid position 665 with alanine (A), substituting glutamine (Q) at the amino acid position 675 with lysine (K), substituting proline (P) at the amino acid position 676 with leucine (L), substituting alanine (A) at the amino acid position 679 with glutamic acid, substituting glutamic acid (E) at the amino acid position 688 with alanine (A), substituting threonine (T) at the amino acid position 716 with alanine (A), substituting alanine (A) at the amino acid position 717 with threonine (T), substituting alanine (A) at the amino acid position 742 with glycine (G), substituting alanine (A) at the amino acid position 783 with valine (V), substituting alanine (A) at the amino acid position 796 with threonine (T), substituting lysine (K) at the amino acid position 814 with glutamic acid (E), substituting isoleucine (I) at the amino acid position 825 with methionine (M), substituting arginine (R) at the amino acid position 826 with serine (S), substituting arginine (R) at the amino acid position 837 with histidine (H), substituting glutamic acid (E) at the amino acid position 871 with asparagine (N), substituting isoleucine (I) at the amino acid position 882 with valine (V), substituting glutamic acid (E) at the amino acid position 888 with glycine (G), substituting aspartic acid (D) at the amino acid position 894 with glycine (G), substituting proline (P) at the amino acid position 895 with serine (S), substituting glycine (G) at the amino acid position 913 with serine (S), substituting glutamic acid (E) at the amino acid position 948 with lysine (K), substituting serine (S) at the amino acid position 955 with asparagine (N), substituting methionine (M) at the amino acid position 968 with valine (V), substituting glutamine (Q) at the amino acid position 971 with glutamic acid (E),) substituting methionine (M) at the amino acid position 980 with valine (V), substituting glutamine (Q) at the amino acid position 982 with arginine (R), substituting alanine (A) at the amino acid position 1009 with aspartic acid (D), substituting aspartic acid (D) at the amino acid position 1020 with glutamic acid (E), substituting histidine (H) at the amino acid position 1022 with tyrosine (Y), substituting glutamine (Q) at the amino acid position 1023 with lysine (K) and glutamic acid (E), and substituting glycine (G) at the amino acid position 1040 with serine (S).

According to the present invention, preparation of the CYP102A1 mutants may be performed using various methods known in the art such as a deletion mutation method (Kowalski D. et al., J. Biochem., 15, 4457), a PCT method, a Kunkel method, a site-directed mutation method, a DNA shuffling, a staggered extension process (StEP), an error-prone polymerase chain reaction (PCR) method, or the like.

According to the present invention, the CYP012A1 mutant may have an amino acid sequence changed from that of the wild-type CYP102A1 by at least one substitution selected from a group consisting of substituting arginine (R) at the amino acid position 47 with leucine (L), substituting tyrosine (Y) at the amino acid position 51 with phenylalanine (F), substituting glutamic acid (E) at the amino acid position 64 with glycine (G), substituting alanine (A) at the amino acid position 74 with glycine (G), substituting phenylalanine (F) at the amino acid position 81 with isoleucine (I), substituting leucine (L) at the amino acid position 86 with isoleucine (I), substituting phenylalanine (F) at the amino acid position 87 with valine (V), substituting glutamic acid (E) at the amino acid position 143 with glycine (G), substituting leucine (L) at

the amino acid position 188 with glutamine (Q), and substituting glutamic acid (E) at the amino acid position 267 with valine (V).

The most preferable CYP102A1 mutant according to the present invention may have an amino acid substitution position and substituted amino acid in the wild-type CYP102A1 selected from a group consisting of F87A, R47L/Y51F, A74G/F87V/188Q, R47L/L86I/L188Q, R47L/F87V/188Q, R47L/F87V/L188Q/E267V, R47L/L86I/F87V/L188Q, R47L/F87V/E143G/L188Q/E267V, R47L/F87V/E143G/L188Q/E267V, and R47L/E64G/F81I/F87V/E143G/L188Q/E267V, and R47L/E64G/F81I/F87V/E143G/L188Q/E267V.

For example, in the CYP102A1 mutant, the amino acid substitution position and substituted amino acid in the wild-type CYP102A1 is F87A, which means that phenylalanine (F) at the amino acid position 87 in the wild-type CYP102A1 is substituted with valine (V). Hereinafter, all of the CYP102A1 mutants and the chimeras derived from the 20 CYP102 A1 mutants may also be interpreted to have the same meaning as described above.

The most preferable chimera derived from the CYP102A1 mutant according to the present invention may have an amino acid substitution position and substituted amino acid in the 25 CYP102A1 mutant selected from a group consisting of A475V/E559D/T665A/P676L/A679E/E688A/A742G/K814E/R826S/R837H/E871N/I882V/E888G/P895S/

S955N/M968V/Q982R/A1009D/H1022Y/Q1023E, A475V/E559D/T665A/A679E/E688A/A742G/K814E/ E871N/I882V/E888G/P895S/G913G/S955N/M968V/ A1009D/H1022Y/Q1023E,

K474T/A475V/A591S/D600É/V625L/D632N/K640A/ T665A/A717T/A742G/A796T/K814E/I825M/I882V/ E888/S955N/M968V/M980V/A1009D/D1020E/ Q1023K/G1040S,

K474T/A475V/R526P/Q547E/D600E/V625 L/D638E/ K640A/G661R/T665A/Q675K/T716A/A717T/A742G/ A783V/K814E/I825M/E871N/I882V/E888G/D894G/ E948K/S955N/M968V/Q971E/A1009D/D1020E,

K474T/A475V/Q513R/Q547E/D600E/V625L/D638E/ K640A/G661R/T665A/Q675K/T716A/A717T/A742G/ A783V/K814E/I825M/E871N/I882V/E888G/D894G/ E948K/S 955N/M968V/A1009D/D1020E,

K474T/A475V/Q547E/D600E/V625L/D638E/K640A/ G661R/T665A/Q675K/T716A/A717T/A742G/A783V/ K814E/I825M/E871N/I882V/E888G/D894G/E948K/ S955N/M968V/A1009D/D1020E, and

K474T/A475V/Q547E/L590F/D600E/V625L/D638E/ K640A/G661R/T665A/Q675K/T716A/A717T/A742G/ A783V/K814E/I825M/E871N/I882V/E888G/D894G/ E948K/S955N/M968V/A1009D/D1020E.

Protein according to the present invention may be prepared using the methods known in the art. For example, protein may be prepared by genetic engineering techniques, peptide synthesis using solid-phase techniques (Merrifield, J. Am. Chem. Soc., 85:2149-2154 (1963)), or method of cleaving protein using peptidase.

Protein according to the present invention may be natural protein or may be prepared by a recombination of culturing 60 cells transformed with DNA encoding CYP102A1 or mutants thereof and collecting the protein. Protein may be prepared by inserting nucleic acid molecules encoding protein according to the present invention into an expression vector, transforming the vector into a host cell, culturing the transformed host cell, and purifying protein expressed by the transformed host cell.

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The vector may be, for example, plasmid, cosmid, a virus, or phage. As the host cell into which DNA in the vector is cloned or expressed, there may be a prokaryotic cell, a yeast cell, and a higher eukaryotic cell. Culture conditions such as a culture medium, a temperature, pH, and the like, may be selected by those skilled in the art without undue experiment. In general, principles, protocols, and techniques for maximizing productivity of the culture of cells may refer to Mammalian Cell Biotechnology: A Practical Approach, M. Butler, ed. (IRL Press, 1991).

The expression and cloning vector may generally include a promoter that is operationally linked to a nucleic acid sequence that encodes CYP102A1 or mutants thereof inducing the synthesis of mRNA. Various promoters that are recognized by host cells are known. A promoter suitable for a prokaryotic host cell may be a β-lactamase and lactose promoter system, alkali phosphatase, a tryptophan (trp) promoter system, and a hybrid promoter, for example, a tac promoter. In addition, the promoter used in bacterial systems may include a Shine-Dalgarno (S.D.) sequence operationally linked to DNA that encodes CYP102A1 mutants. An example of the promoter suitable for a yeast host cell may include 3-phosphooglycerate kinase or other glycosidase.

The method for preparing 2-hydroxyatorvastin or 4-hydroxylated product from atorvastatin according to the present invention may further include adding a NADPH-generating system.

The NADPH-generating system may include glucose 6-phosphate, NADP+, and yeast glucose 6-phosphate dehydrogenase, but is not limited thereto.

In the NADPH-generating system, in the case in which the wild-type CYP102A1, the CYP012A1 mutants, and the chimeras derived from the CYP102A1 mutants are reacted with atorvastatin together with each other, atorvastatin may be effectively converted into 2-hydroxylated product and 4-hydroxylated product at the same time.

In addition, the method for preparing 2-hydroxylated product or 4-hydroxylated product from atorvastatin according to the present invention may be performed at 0 to 40° C., and preferably, 30 to 40° C. At the time of oxidation reaction using atorvastatin as the substrate in vitro system, the catalytic activity is increased at this temperature, thereby making it 45 possible to efficiently and selectively produce atorvastatin.

In another general aspect, the present invention provides a kit for preparing 2-hydroxylated product or 4-hydroxylated product from atorvastatin including at least one enzyme selected from a group consisting of the wild-type CYP102A1, the CYP102A1 mutants, and the chimeras derived from the CYP102A1 mutants and the NADPH-generating system,

wherein the CYP102A1 mutant includes an amino acid substitution position and substituted amino acid in the wild-type CYP102A1 selected from a group consisting of F87A, R47L/Y51F, A74G/F87V/L188Q, R47L/L86I/L188Q, R47L/F87V/L188Q, R47L/L86I/L188Q/E267V, R47L/L86I/L188Q/E267V, R47L/L86I/L188Q/E267V, R47L/L86I/F87V/L188Q, R47L/F87V/E143G/L188Q/E267V, R47L/F81I/F87V/E143G/L188Q/E267V, and R47L/E64G/F81I/F87V/E143G/L188Q/E267V, and

the chimera derived from the CYP102A1 mutant includes an amino acid substitution position and substituted amino acid in the CYP102A1 mutant selected from a group consisting of

65 A475V/E559D/T665A/P676L/A679E/E688A/A742G/ K814E/R826S/R837H/E871N/I882V/E888G/P895S/ S955N/M968V/Q982R/A1009D/H1022Y/Q1023E,

- A475V/E559D/T665A/A679E/E688A/A742G/K814E/ E871N/I882V/E888G/P895S/G913G/S955N/M968V/ A1009D/H1022Y/Q1023E,
- K474T/A475V/A591S/D600E/V625L/D632N/K640A/ T665A/A717T/A742G/A796T/K814E/I825M/I882V/ E888/S955N/M968V/M980V/A1009D/D1020E/ Q1023K/G1040S,
- K474T/A475V/R526P/Q547E/D600E/V625L/D638E/ K640A/G661R/T665A/Q675K/T716A/A717T/A742G/ A783V/K814E/I825M/E871N/I882V/E888G/D894G/ E948K/S955N/M968V/Q971E/A1009D/D1020E,
- K474T/A475V/Q513R/Q547E/D600E/V625L/D638E/ K640A/G661R/T665A/Q675K/T716A/A717T/A742G/ A783V/K814E/I825M/E871N/I882V/E888G/D894G/ E948K/S955N/M968V/A1009D/D1020E,
- K474T/A475V/Q547E/D600V625L/D638E/K640A/ G661R/T665A/Q675K/T716A/A717T/A742G/A783V/ K814E/I825M/E871N/I882V/E888G/D894G/E948K/ S955N/M968V/A1009D/D1020E, and
- K474T/A475V/Q547E/L590F/D600E/V625L/D638E/ K640A/G661R/T665A/Q675K/T716A/A717T/A742G/ A783V/K814E/I825M/E871N/I882V/E888G/D894G/ E948K/S955N/M968V/A1009D/D1020E.

The kit according to the present invention may further include a reagent required to progress the reaction.

The NADPH-generating system may include glucose 6-phosphate, NADP+, and yeast glucose 6-phosphate dehydrogenase, but is not limited thereto.

#### Advantageous Effects of Invention

As set forth above, the wild-type CYP102A1, the CYP102A1 mutants, and the chimeras derived from the CYP102A1 mutants according to the present invention may stably and efficiently serve as the catalyst in the reaction of 35 converting atorvastatin into 2-hydroxylated product and 4-hydroxylated product, such that 2-hydroxylated product and 4-hydroxylated product may be environmentally-friendly and selectively prepared on a large scale.

The composition, the kit, and the method for preparing 40 2-hydroxylated product or 4-hydroxylated product according to the present invention includes the wild-type CYP102A1, the CYP102A1 mutants, or the chimeras derived from the CYP102A1 mutants, such that 2-hydroxylated product or 4-hydroxylated product may be economically, efficiently, and 45 selectively prepared from atorvastatin on a large scale. Therefore, the present invention may contribute to developing novel drugs using the metabolites of atorvastatin.

#### BRIEF DESCRIPTION OF DRAWINGS

The above and other objects, features and advantages of the present invention will become apparent from the following description of preferred embodiments given in conjunction with the accompanying drawings, in which:

FIG. 1 shows an amino acid sequence (sequence No. 16) of a wild-type CYP102A1 according to an exemplary embodiment of the present invention;

FIG. 2 shows a nucleotide sequence (sequence No. 17) of a wild-type CYP102A1 according to another exemplary 60 embodiment of the present invention;

FIG. 3 shows an amino acid sequence (sequence No. 18) of a wild-type CYP102A1 mutant #16 according to another exemplary embodiment of the present invention;

FIG. 4 shows a nucleotide sequence (sequence No. 19) of a 65 wild-type CYP102A1 mutant #16 according to another exemplary embodiment of the present invention;

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FIG. 5 shows an amino acid sequence (sequence No. 20) of a wild-type CYP102A1 mutant #17 according to another exemplary embodiment of the present invention;

FIG. 6 shows a nucleotide sequence (sequence No. 21 of a wild-type CYP102A1 mutant #17 according to another exemplary embodiment of the present invention;

FIG. 7 shows an amino acid sequence (sequence No. 22) of a chimera #16A1V2 derived from the wild-type CYP102A1 mutant #16 according to another exemplary embodiment of the present invention;

FIG. 8 shows a nucleotide sequence (sequence No. 23) of a chimera #16A1V2 derived from the wild-type CYP102A1 mutant #16 according to another exemplary embodiment of the present invention;

FIG. **9** shows high-performance liquid chromatography (HPLC) chromatograms (measuring UV absorbance at 260 nm) of atorvastatin metabolites produced by human CYP3A4;

FIGS. **10**A and **10**B show high-performance liquid chromatography (HPLC) chromatograms (measuring UV absorbance at 260 nm) of atorvastatin metabolites produced by a CYP102A1 mutant (FIG. **10**A) and a chimera (FIG. **10**B) derived from a CYP102A1 mutant according to the exemplary embodiment of the present invention;

FIGS. 11A and 11B show LC-MS elution profiles of atorvastatin and metabolites thereof produced by the human CYP3A4 (FIG. 11A) and the chimera #16A1V2 derived from the CYP102A1 mutant according to the exemplary embodiment of the present invention (FIG. 11B);

FIGS. **12**A to **12**C show LC-MS elution profiles of atorvastatin and metabolites thereof produced by a chimera (#16A1V2) derived from the CYP102A1 mutant according to the exemplary embodiment of the present invention;

(A: 4-hydroxylated product, B: 2-hydroxylated product, C: atorvastatin)

FIG. 13 shows turnover numbers of atorvastatin oxidation using the wild-type CYP102A1, mutants and the chimera derived from the CYP102A1 mutants according to the exemplary embodiment of the present invention; and

FIG. 14 shows total turnover numbers (TTNs) of atorvastatin oxidation using chimeras derived from specific CYP102A1 mutants according to the exemplary embodiment of the present invention.

#### MODE FOR THE INVENTION

Hereinafter, exemplary embodiments of the present invention will be described in detail with reference to the accompanying drawings so that those skilled in the art may easily practice the present invention. However, the embodiment of the present invention has been disclosed for illustrative purposes, but the scopes of the present invention are not limited thereby.

#### EXAMPLE 1

### Construction of P450 BM3 Mutants by Site-directed Mutagenesis

17 site-directed mutants of CYP102A1 were prepared by the same method as a method used by Kim et al., (Drug Metab. Dispos. 35: 2166-2170, 2008b). Primers used in order to introduce BanHI/SacI restriction sites and polymerase chain reaction (PCR) primers in order to introduce mutation were shown in the following Table 1. Codons for amino acid substitution were in italics and are underlined. The PCR primers were obtained from Genotech (Daejeon, Korea). Genes

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encoding the CYP102A1 mutants were amplified from pCWBM3 by PCR primers designed to facilitate cloning into an expression vector pCWori (Dr. F. W. Dahlquist, University of California, Santa Barbara, Calif.) or pSE420 (Invitrogen).

Oligonucleotide assembly was performed using the 14 sets of the designed primers shown in the following Table 1. The amplified genes were cloned into the BanHI/SacI restriction sites of the PCWBM3 BanHI/SacI vector. These plasmids were transformed into *Escherichia coli* DH5αF-IQ (Invitrogen), and this strain was also used to express the mutant CYP102A1 proteins. After mutagenesis, whether or not the desired mutations were generated was confirmed by DNA sequencing (Genotech, Daejeon, Korea).

TABLE 1

	Prim	ers used	to prepare mutants
Name			Sequence
BamHI forv		1)	5' -AGC GGA TCC ATG ACA ATT AAA GAA ATG CCT C-3'
SacI revenue (sequence		2)	5' -ATC GAG CTC GTA GTT TGT AT-3'
R47L (sequence	list	3)	5' -GCG CCT GGT <u>CTG</u> GTA ACG CG-3'
Y51F (sequence	1ist	4)	5' -GTA ACG CGC <u>TTC</u> TTA TCA AGT-3'
E64G (sequence	list	5)	5' -GCA TGC GAT <u>GGC</u> TCA CGC TTT-3'
A74G (sequence	list	6)	5' -TA AGT CAA <u>GGC</u> CTT AAA TTT GTA CG-3'
F81I (sequence	list	7)	5' -GTA CGT GAT <u>ATT</u> GCA GGA GAC-3'
L861 (sequence	list	8)	5' -GGA GAC GGG <u>ATT</u> TTT ACA AGC T-3'
F87A (sequence	list	9)	5' -GAC GGG TTA <u>GCG</u> ACA AGC TGG-3'
F87V (sequence	list	10)	5' -GAC GGG TTA <u>GTG</u> ACA AGC TGG-3'
E143G (sequence	list	11)	5' -GAA GTA CCG <u>GGC</u> GAC ATG ACA-3'
L188Q (sequence	list	12)	5'-ATG AAC AAG <u>CAG</u> CAG CGA GCA A-3'
A264G (sequence	list	13)	5' -TTC TTA ATT <u>GGG</u> GGA CAC GTG-3'
E267V (sequence	list	14)	5' -T GCG GGA CAC <u>GTG</u> ACA ACA AGT-3'

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TABLE 1-continued

	Primers used	l to prepare mutants
Name		Sequence
/		
L861/F87V (sequence	list 15)	5' -GGA GAC GGG <u>ATT</u> <u>GTG</u> ACA AGC TG-3'

#### EXAMPLE 2

Expression and Purification of Wild-type CYP102A1, Wild-type CYP102A1 Mutants, and Chimeras Derived from CYP012A1 Mutant

Plasmids including genes of the Wild-type CYP102A1 (pCWBM3) and CYP102A1 mutant were transformed into Escherichia coli DH5αF-IQ (Kim et al., Drug Metab. Dispos. 35:2166-2170, 2008b). A culture was inoculated from a 20 single colony into 5 ml of a Luria-Bertani medium supplemented with ampicillin (100 μg/ml) and grown at 37° C. This culture was inoculated into 250 ml of a Terrific Broth medium supplemented with ampicillin (100 μg/ml) and grown at 37° C. with shaking at 250 rpm so as to reach OD600 of about 0.8, and then gene expression was induced by the addition of isopropyl-β-D-thiogalactopyranoside to a final concentration of 0.5 mM. δ-Aminolevulinic acid (0.1 mM) was also added thereto. After inducing the expression, the culture was allowed to grow another 36 hours at 30° C., and then cells were harvested by centrifugation (15 min, 5000 g, 4° C.). The cell pellet was resuspended in a TES buffer solution (100 mM Tris-HCL, pH 7.6, 500 mM sucrose, 0.5 mM EDTA) and lysed by sonication (Sonicator; Misonix, Inc., Farmingdale. N.Y.). After the lysates was centrifuged at 100,000 g (90 min. 4° C.), a soluble cytosolic fraction was collected and used for 35 the activity assay. The soluble cytosolic fraction was dialyzed from a 50 mM potassium phosphate buffer (pH 7.4) and stored at -80° C. The cytosolic fraction was used within 1 month of manufacture.

The CYP102A1 concentrations were determined from CO-difference spectra using  $\epsilon$ =91 mM/cm (Omura and Sato. J. Biol. Chem. 239:2370-2378, 1964). For all of the wild-types and mutants, a typical culture yielded 300 to 700 nM P450. The expression level of wild-type CYP102A1 and the mutants thereof were in the range of 1.0 to 2.0 nmol P450/mg cytosolic protein.

Several mutants with high catalytic activity for some substrates in human were selected among the prepared mutants, and the amino acid substitution sites in the mutants were shown in Tables 2 and 3.

[References]

Carmichael and Wong. Eur. J. Biochem. 268:3117-3125, 2001; Li et al., Appl. Environ. Microbiol. 67:5735-5739, 2001; van Vugt-Lussenburg et al., J. Med. Chem. 50:455-461, 2007

TABLE 2

CYP102A1 mutants used in the present invention								
Abbreviations	BM3 wild type and mutants	Ref						
WT	BM3 wild type	Carmichael and Wong, 2001						
Mutant #1	F87A	Carmichael and Wong, 2001						
Mutant #2	A264G	Carmichael and Wong, 2001						
Mutant #3	F87A/A264G	Carmichael and Wong, 2001						
Mutant #4	R47L/Y51F	Carmichael and Wong, 2001						
Mutant #5	R47L/Y51F/A264G	Carmichael and Wong, 2001						
Mutant #6	R47L/Y51F/F87A	Carmichael and Wong, 2001						
Mutant #7	R47L/Y51F/F87A/A264G	Carmichael and Wong, 2001						
Mutant #8	A74G/F87V/L188Q	Li et al., 2001						

TABLE 2-continued

	CYP102A1 mutants used in the present invention								
Abbreviations	BM3 wild type and mutants	Ref							
Mutant #9	R47L/L86I/L188Q	Kim et al., 2008b							
Mutant #10	R47L/F87V/L188Q	van Vugt-Lussenburg et al., 2007							
Mutant #11	R47L/F87V/L188Q/E267V	van Vugt-Lussenburg et al., 2007							
Mutant #12	R47L/L86I/L188Q/E267V	Kim et al., 2008b							
Mutant #13	R47L/L86I/F87V/L188Q	van Vugt-Lussenburg et al., 2007							
Mutant #14	R47L/F87V/E143G/L188Q/E267V	Kim et al., 2008b							
Mutant #15	R47L/E64G/F87V/E143G/L188Q/E267V	Kim et al., 2008b							
Mutant #16	R47L/F81I/F87V/E143G/L188Q/E267V	Kim et al., 2008b							
Mutant #17	R47L/E64G/F81I/F87V/E143G/L188Q/E267	7V van Vugt-Lussenburg et al., 2007							

TABLE 3

	Mutated Amino acid	Change of Nucleotide	*2	*3	*4	*5	*6	*7	*8	*9	QMB1551
	T2P	4A > C									+
Heme	V27I	79G > A	+		+		+	+	+	+	+
domain	A29T	85G > A	+		+		+	+	+	+	+
	V128I	382G > A	+		+	+	+	+	+	+	+
	A136T	406G > A	+		+		+	+	+	+	+
	E208D	624A > C				+					
	A222T	664G > A									+
	A296T	886G > A	+		+						
	D370E	1110C > A	+		+						
	K453Q	1357A > C				+	+	+	+	+	+
	T464R	1392T > A				+	+	+	+	+	+
	V471E	1413A > G				+	+	+	+	+	+
Reductase	K474T	1422G > C				+	+	+	+	+	+
domain	A475V	1424C > T	+	+	+	+	+	+	+	+	+
	Q513R	1539G > A						+			
	R526P	1578C > T					+				
	Q547E	1639C > G					+	+	+	+	+
	E559D	1677A > C	+	+	+						
	L590F	1794C > A								+	
	A591S	1771G > T				+					
	D600E	1800C > A				+	+	+	+	+	+
	V625L	1873G > T				+	+	+	+	+	+
	D632N	1894G > A				+					
	D638E	1914T > A					+	+	+	+	+
	K640A	1920A > T				+	+	+	+	+	+
	A652S	1954G > T									+
	G661R	1981G > C					+	+	+	+	+
	T665A	1993A > G	+	+	+	+	+	+	+	+	+
	Q675K	2023C > A					+	+	+	+	+
	P676L	2027C > T	+	+							
	A679E	2036C > A	+	+	+						
	E688A	2063A > C	+	+	+						
	T716A	2146A > G					+	+	+	+	+
	A717T	2149G > A				+	+	+	+	+	+
	A742G	2225C > G	+	+	+	+	+	+	+	+	+
	A783V	2348C > T					+	+	+	+	+
	A796T	2386G > A				+					
	K814E	2440A > G	+	+	+	+	+	+	+	+	+
	I825M	2474A > G				+	+	+	+	+	+
	R826S	2476C > A	+	+							
	R837H	2510G > A	+	+							
	E871N	2613G > T	+	+	+		+	+	+	+	+
	I882V	2644A > G	+	+	+	+	+	+	+	+	+
	E888G	2663A > G	+	+	+	+	+	+	+	+	+
	D894G	2681A > G					+	+	+	+	+
	P895S	2683C > T	+	+	+						
	G913S	2739C > T			+						
	E948K	2842G > A					+	+	+	+	+
	S955N	2864G > A	+	+	+	+	+	+	+	+	+
	M968V	2904G > A	+	+	+	+	+	+	+	+	+
	Q971E	2911C > G					+				
	M980V	2938A > G				+					
	Q982R	2945A > G	+	+							
	A1009D	3026C > A	+	+	+	+	+	+	+	+	+

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TABLE 3-continued

CYP102A1 natural variants used in the present invention CYP102A1 Variants										
Mutated Amino acid	Change of Nucleotide	*2	*3	*4	*5	*6	*7	*8	*9	QMB1551
H1022Y	3066C > T	+	+	+						
Q1023K	3067C > G				+					
Q1023E	3067C > A	+	+	+						
G1040S	3118G > A				+					

In addition, a chimeric protein of selective CYP102A1 mutants was constructed by fusing heme domains of the prepared CYP102A1 mutants of Tables 2 and 3 to reductase domains of the natural variants of the wild-type CYP102A1.

In order to clone the chimeric protein of the selective CYP102A1 mutant prepared by fusing the heme domain and the reductase domain to each other, the chimeric protein was cloned into the expression vector pCW vector prepared using <sup>20</sup> BanHI/SacI and SacI/XhoI.

Plasmids including genes of the chimeric protein of the CYP102A1 mutant were transformed into *Escherichia coli* DH5 $\alpha$ F-IQ (Kim et al. Protein Expr. Purif. 57:188-200, 2008). A culture was inoculated from a single colony into 5 ml of a Luria-Bertani medium supplemented with ampicillin (100 µg/ml) and grown at 37° C. This culture was inoculated into 250 ml of a Terrific Broth medium supplemented with ampicillin (100 µg/ml) and grown at 37° C. with shaking at 250 rpm so as to reach OD600 of about 0.8, and then gene expression was induced by the addition of isopropyl- $\beta$ -D-thiogalactopyranoside to a final concentration of 0.5 mM.

δ-Aminolevulinic acid (0.1 mM) was also added thereto. After inducing of the expression, the culture was allowed to grow another 36 hours at 30° C., and then cells were harvested by centrifugation (15 min, 5000 g, 4° C.). The cell pellet was resuspended in a TES buffer solution (100 mM Tris-HCL, pH 7.6, 500 mM sucrose, 0.5 mM EDTA) and lysed by sonication

(Sonicator. Misonix. Inc., Farmingdale. N.Y.). After the lysates was centrifuged at 100,000 g (90 min, 4° C.), a soluble cytosolic fraction was collected and used for the activity assay. The soluble cytosolic fraction was dialyzed from a 50 mM potassium phosphate buffer (pH 7.4) and stored at -80° C. The cytosolic fraction was used within 1 month of manufacture.

The CYP102A1 concentrations were determined from CO-difference spectra using  $\epsilon$ =91 mM/cm (Omura and Sato, J. Biol. Chem. 239:2379-2385 1964). For the chimeras derived from CYP102A1, a typical culture yielded 300 to 700 nM P450. The expression levels of the chimeras derived from the CYP102A1 mutant were in the range of 1.0 to 2.0 nmol P450/mg cytosolic protein.

Several chimeras with high catalytic activity for some substrates in a human were selected among the chimeras prepared from the CYP102A1 mutants, and the amino acid substitution sites in each chimera were shown in Table 4 (Kang et al., AMB Express, 1:1, 2011).

Hereinafter, the chimeras derived from the CYP102A1 mutants used in this experiment were called as follows.

In the present invention, the terms chimera #16A1V2 of the mutants means a chimera derived from a CYP102A1 mutant #16 prepared by fusing the heme domains of the mutant #16 in Table 2 to V2 reductase domain of the following Table 4.

TABLE 4

	CYP102A1 natural variants used in the present invention	
Abbreviations	Natural variants	Ref
variant2(V2)	A475V/E559D/T665A/P676L/A679E/E688A/A742G/K814E/ R826S/R837H/E871N/I882V/E888G/P895S/S955N/M968V/ O982R/A1009D/H1022Y/O1023E	Kang et al. 2011
variant3(V3)	A475V/E559D/T665A/P676L/A679E/E688A/A742G/K814E/ R826S/R837H/E871N/I882V/E888G/P895S/S955N/M968V/ Q982R/A1009D/H1022Y/Q1023E	Kang et al. 2011
variant4(V4)	A475V/E559D/T665A/A679E/E688A/A742G/K814E/E871N/ 1882V/E888G/P895S/G913G/S955N/M968V/A1009D/H1022Y O1023E	Kang et al. 2011
variant5(V5)	K474T/A475V/A591S/D600E/V625L/D632N/K640A/T665A/ A717T/A742G/A796T/K814E/I825M/I882V/E888/S955N/ M968V/M980V/A1009D/D1020E/Q1023E/G1040S	Kang et al. 2011
variant6(V6)	K474T/A475V/R526P/Q547E/D600E/V625L/D638E/K640A/ G661R/T665A/Q675K/T71GA/A717T/A742G/A783V/K814E/ I825M/E871N/I882V/E888G/D894G/E948K/S955N/M968V/ Q971E/A1009D/D1020E	Kang et al. 2011
variant7(V7)	K474T/A475V/Q513R/Q547E/D600E/V625L/D638E/K640A/ G661R/T665A/Q675K/T716A/A717T/A742G/A783V/K814E/ I825M/E871N/I882V/E888G/D894G/E948K/S955N/M968V/ A1009D/D1020E	Kang et al. 2011
variant8(V8)	K474T/A475V/Q547E/D600E/V625L/D638E/K640A/G661R/ T665A/Q675K/T716A/A717T/A742G/A783V/K814E/I825M/ E87IN/I882V/E888G/D894G/E948K/S955N/M968V/A1009D/ D1020E	Kang et al. 2011
variant9(V9)	K474T/A475V/Q547E/L590F/D600E/V625L/D638E/K640A/ G661R/T665A/Q675K/T716A/A717T/A742G/A783V/K814E/ I825M/E871N/I882V/R888G/D894G/E948K/S955N/M968V/ A1009D/D1U20E	Kang et al. 2011

#### **EXAMPLE 3**

Oxidation of Atorvastatin by Wild-type CYP102A1, Wild-type CYP102A1 Mutants, and Chimeras Derived from CYP102A1 Mutant

It was examined whether the wild-type CYP102A1, the CYP102A1 mutants, and the chimeras derived from the CYP1.02A1 mutants may oxidize atorvastatin. Typical steady-state reactions was performed by adding 50 pmol CYP102A1 and 80 μM substrate to 0.25 ml of 100 mM potassium phosphate buffer solution (pH 7.4). In order to initiate reactions, the NADPH-generating system was added thereto (final concentrations: 10 mM glucose 6-phosphate, 0.5 mM NADP+, and 1 IU yeast glucose 6-phosphate per ml). A stock solution of atorvastatin (20 mM) was prepared in DMSO and diluted into the enzyme reaction solution to have a final organic solvent concentration of <1% (v/v).

In order to measure human CYP3A4 activity, 50 pmol P450, 100 pmol NADPH-P450 reductase (CPR), 100 pmol  $^{20}$  cytochrome b5, and  $^{45}$   $\mu$ M L- $\alpha$ -dilauroyl-sn-glycero-3-phosphocholine (DLPC) were used instead of 50 pmol CYP102A1. After the reaction solution was reacted for 30 minutes at  $^{30}$  C., the reaction was terminated with 2-fold of ice-cold dichloromethane.

#### (1) HPLC Analysis

After centrifugation of the reaction mixture, a supernatant was removed and a solvent was evaporated under nitrogen gas and analyzed using HPLC. A sample (30 ul) was injected into Gemini C18 column (4.6 mm×150 mm, 5 um. Phenomenex.

Torrance, Calif.). As a mobile phase A, water containing 0.1% formic acid/acetonitrile (80/20, v/v) was used, and as a mobile phase B, acetonitrile/0.1% formic acid (90/10, v/v) was used. The mobile phase A/B (70/30, v/v) was flowed at a rate of 1 ml·min<sup>-1</sup> using a gradient pump (LC-20AD, Shimadzu, Kyoto, Japan). Elution solutions were detected by UV at 260 nm.

In order to examine whether or not CYP102A1 (P450 BM3) may oxidize atorvastatin, the abilities of the wild-type CYP102A1 (P450 BM3), the mutants thereof, and the chimeas derived from the CYP102A1 mutants to oxidize atorvastatin were measured at a fixed substrate concentration (80  $\mu$ M).

The metabolites of atorvastatin prepared by the human CYP3A4, the bacterial CYP102A1 mutant (#16 in Table 2), 45 and the chimera (#16A1V3) derived from the CYP102A1 were examined using HPLC chromatograms (measuring UV absorbance at 260 nm).

Peaks were confirmed by comparing with retention times of peaks of the metabolites prepared by human CYP3A4 and 50 CYP2C9. The substrate and two main metabolites, that is, 2-hydroxylated product and 4-hydroxylated product were shown

As a result, it might be appreciated that retention times of the peaks of the metabolites exactly coincide with those of the 55 standard 4-OH atorvastatin and 2-OH atorvastatin as shown in FIGS. 9 to 10B.

#### (2) LC-MS Analysis

In order to identify atorvastatin metabolites produced the wild-type CYP102A1 mutants and the chimeras derived from 60 by CYP102A1 mutants, LC-MS analysis was conducted by comparing LC profiles and fragmentation patterns of atorvastatin and metabolites thereof.

The wild-type CYP102A1 mutants and human CYP3A4 were incubated with 80  $\mu M$  of atorvastatin at 37° C. for 30 minutes in the presence of an NADPH-generating system. Reactions were terminated by the addition of 2-fold ice-cold

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 $CH_2Cl_2.$  After centrifugation of the reaction mixture, a supernatant was removed and an organic solvent layer was evaporated under nitrogen. The reactant was reconstituted into 100  $\mu l$  of a mobile phase by vortex mixing and sonication for 20 sec. An aliquot (10  $\mu l)$  of the prepared solution was injected into the LC column.

LC-MS analysis was carried out on Shimadzu LCMS-2010 EV system (Shimadzu Corporation, Japan) having LCMS solution software by electro spray ionization in a positive mode. In a Shim-pack VP-ODS column (250 mm×2.0 mm i.d., Shimadzu Corporation, Japan) water containing 0.1% formic acid/acetonitrile (80/20, v/v) was used as a mobile phase A, and acetonitrile/0.1% formic acid (90/10, v/v) was used as a mobile phase B. The mobile phase A/B (70/30, v/v)was separated using a gradient pump (LC-20AD, Shimadzu. Kyoto, Japan) at a flow rate of 0.16 ml/min. In order to identify the metabolites, mass spectra were recorded by electro spray ionization in a negative mode. Interface and detector voltages are 4.4 kV and 1.5 kV, respectively. Nebulization gas flow was set at 1.5 ml/min. and interface, curve desolvation line (CDL), and heat block temperatures were 250, 230, and 200° C., respectively.

As a result, it might be appreciated that in mass spectra of the reaction samples, peaks were observed at 7.183 min (4-OH atorvastatin), 19.583 min (2-OH atorvastatin), and 21.450 min (atorvastatin) as shown in total ion current (TIC) profiles of the metabolites prepared by the human CYP3A4 (A) and the chimera #16A1V2 (B) derived from the CYP102A1 mutant of FIG. 11.

Further, as shown in FIGS. **12**A to **12**C, the peaks in mass spectra of 4-hydroxylated products (A), 2-hydroxylated products (B), and atorvastatin products (C) by the chimera #16A1V2 derived from the CYP102A1 mutant were observed at 573, 573, and 557, respectively, when calculated as [M-H]<sup>-</sup>.

Based on the results of LC-MS analysis of the reactants, it might be appreciated that the CYP102A1 mutants and the chimeras derived from the CYP102A1 mutants produce 4-hydroxylated or 2-hydroxylated product from atorvastatin. The retention time and fragmentation pattern of the metabolites produced by the CYP102A1 mutants and the chimeras derived from the CYP102A1 mutants were exactly matched to those of authentic metabolites produced by human CYP3A4.

#### (3) Determination of Turnover Number

In order to recognize production rate of atorvastatin oxides by wild-type CYP102A1, CYP102A1 mutants, and chimeras derived from the CYP102A1 mutants, the turnover number was determined in the reaction using  $80~\mu M$  statin.

The term "turnover number" means the number of substrate molecules that a molecule of an enzyme may convert into products per minute and indicates conversion frequency.

The production rate of 4-hydroxylated metabolite was determined by HPLC as described above.

As shown in FIG. 13, it might be appreciated that three kinds of mutants (#15, #16, and #17 in Table 2) and five kinds of chimeras (#16A1V2, #16A1V3, #17A1V2, #17A1V3, and #17A1V8) derived from the mutants have high turnover number as the results of measuring the turnover numbers of 17 kinds of mutants and 7 kinds of chimeras derived from the mutants in oxidation of atorvastatin (producing the metabolites of atorvastatin).

Particularly, it might be appreciated that the chimeras #16A1V2 and #17A1V2 derived from the mutants have the same activity as that of the human CYP3A4.

In order to recognize production rate of atorvastatin metabolites by the CYP102A1 mutant (#16 in Table 2) and

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the chimeras (#16A1V2 and #17A1V2) derived from the CYP102A1 mutants, total turnover numbers (TTNs; mol product/mol catalyst) were determined in reactions using total 240 uM atorvastatin.

The term "total turnover number (TTN)" means the number of substrate molecules converted into metabolites by enzymes for the total reaction time.

The total turnover numbers (TTNs) were determined by comparing the results under three conditions. First, the reaction was performed by adding a NADPH-generating system at 37° C. for 1 hour in the presence of 80 µM substrate. In addition, second, after reaction was performed for 1 hour in the presence of 80 µM substrate, 80 µM substrate was additionally added to the reaction mixture, and the reaction was further performed for 1 hour. Finally, after reaction was performed for 1 hour in the presence of 80 µM substrate, 80 µM substrate was additionally added to the reaction mixture, and the reaction was further performed for 1 hour. Then, 80 µM substrate was additionally added to the reaction mixture, and 20 the reaction was further performed for 2 hours.

The production rate of the atorvastatin metabolites was determined using HPLC. The enzyme capable of most efficiently producing a large amount of metabolites in vitro may be selected by comparing the results according to concentra- 25 tion of the substrate and reaction time using mutants or chimeras derived from the mutants having higher activity based on experimental results of the turnover number.

As a result, the total turnover numbers (TTNs; mol product/mol catalyst) were in a range of 31 to 83 as shown in FIG.

Particularly, when the chimeras #16A1V2 and #17A1V2 derived from CYP102A1 mutants having high activity were reacted for 4 hours, it might be appreciated that #16A1V2 has activity higher than that of the human CYP3A4.

The production of metabolites of atorvastatin by chemical synthesis has never been reported up to now. Therefore, it may be an alternative to chemical synthesis of the target metabolites in the Examples of the present invention to use CYP102A1 enzymes, that is, CYP102A1 mutants and the

<160> NUMBER OF SEQ ID NOS: 23

chimeras derived from the CYP102A1 mutants to generate the metabolites of atorvastatin according to the present inven-

According to the present invention, it might be appreciated that bacterial CYP102A1 enzymes of the Examples catalyze the same reaction as that of the human CYP3A4 to produce 4-OH product and 2-OH product, which are the human metabolites.

In addition, it might be appreciated that the wild-type CYP102A1 mutants and the chimeras derived from the CYP102A1 mutants catalyze oxidation of atorvastatin, which is the human P450 substrate, and produces 4-hydroxylated product and 2-hydroxylated product, which are the main metabolites produced by the human CYP3A4, from atorvas-

Further, it may be appreciated that the wild-type CYP102A1 mutants and the chimeras derived from the CYP102A1 mutants according to the present invention may efficiently produce the human metabolites from atorvastatin, these metabolites may be used to estimate effect, toxicity, and pharmacokinetics of drugs, or the like in a process of developing the drugs, and used to prepare human metabolite derivatives capable of serving as a lead compound of developing the drug.

Sequence Listing Free Text

SEQ. ID. NO: 1 to 15 are primer sequence

SEQ. ID. NO: 16 is an amino acid sequence of a wild-type CYP102A1

SEQ. ID. NO: 17 is a nucleotide sequence of a wild-type CYP102A1

SEQ. ID. NO: 18 is an amino acid sequence of a wild-type CYP102A1 mutant #16

SEQ. ID. NO: 19 is a nucleotide sequence of a wild-type CYP102A1 mutant #16

SEQ. ID. NO: 20 is an amino acid sequence of a wild-type CYP102A1 mutant #17

SEQ. ID. NO: 21 is a nucleotide sequence of a wild-type CYP102A1 mutant #17

SEQ. ID. NO: 22 is an amino acid sequence of a chimera #16A1V2 derived from the wild-type CYP102A1 mutant #16 SEQ. ID. NO: 23 is a nucleotide sequence of a chimera #16A1V2 derived from the wild-type CYP102A1.

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20 25 30  Ile Ala Asp Glu Leu Gly Glu Ile Phe Lys Phe Glu Ala Pro Gly Arg 45  Val Thr Arg Tyr Leu Ser Ser Gln Arg Leu Ile Lys Glu Ala Cys Asp 50  Glu Ser Arg Phe Asp Lys Asn Leu Ser Gln Ala Leu Lys Phe Val Arg 80  Asp Phe Ala Gly Asp Gly Leu Phe Thr Ser Trp Thr His Glu Lys Asn 95  Trp Lys Lys Ala His Asn Ile Leu Leu Pro Ser Phe Ser Gln Gln Ala
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So S
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85 90 95  Trp Lys Lys Ala His Asn Ile Leu Leu Pro Ser Phe Ser Gln Gln Ala
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Gly Lys Asp Pro Glu Thr Gly Glu Pro Leu Asp Asp Glu Asn Ile Arg

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<sup>&</sup>lt;213 > ORGANISM: Artificial sequence

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The invention claimed is:

A method for preparing a 2-hydroxylated product or a <sup>30</sup>
 4-hydroxylated product of atorvastatin comprising:

reacting at least one enzyme comprising a chimera derived from a CYP102A1 mutant with atorvastatin for a time period of at least two hours,

wherein the CYP102A1 mutant has an amino acid <sup>35</sup> sequence changed from that of the wild-type CYP102A1 by at least one substitution selected from the group consisting of R47L/E64G/F87V/E143G/L188Q/E267V, R47L/F81I/F87V/E143G/L188Q/E267V, and R47L/E64G/F81I/F87V/E143G/L188Q/E267V and 40 wherein the chimera from the CYP102A1 mutant includes an amino acid substitution position and substituted amino acid in the CYP102A1 mutant selected from the group consisting of

A475V/E559D/T665A/P676L/A679E/E688A/A742G/ K814E/R826S/R837H/E871N/1882V/E88 8G/P895S/ S955N/M968V/Q982R/A1009D/H1022Y/Q1023E, A475V/E559D/T665A/A679E/E688A/A742G/K814E/ E871N/1882V/E888G/P895S/G913G/S95 5N/M968V/ A1009D/H1022Y/Q1023E, and

K474T/A475V/Q547E/D600EN625L/D**638**E/K640A/G661R/T665A/Q675K/T716A/A717T/A7 42G/A783V/K814E/1825M/E871N/1882V/E888G/D894G/E948K/S955N/M968V/A1009D/D10 20E; and

wherein total turnover number of atorvastatin is increased as compared to same reaction in which the CYP102A1 mutant is reacted with atorvastatin.

- **2**. The method of claim **1**, further comprising adding a NADPH-generating system.
- 3. The method of claim 2, wherein the NADPH-generating system includes glucose 6-phosphate, NADP+, and yeast glucose 6-phosphate dehydrogenase.

\* \* \* \* \*